

Jake Ireland

Mutations and Human Health

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Module Synopsis

The module provides an advanced understanding of the effects of carcinogenic agents on human health, and develops skills in investigating and assessing DNA damage caused by genotoxic compounds to improve the prevention and treatment of cancer and human disease.

Learning Outcomes

Demonstrate a critical understanding of carcinogenic compounds, and their uses and existence in a historical context.

Demonstrate a critical understanding of the mode of action of DNA damaging agents with respect to their carcinogenicity, and be able to critically discuss exposure through to cancer.

Evaluate genetic toxicological data in a qualitative and quantitative manner.

Demonstrate critical knowledge of the current genotoxic compounds in the public eye, and be able to have an unbiased view on the potential carcinogenicity of these agents.

Implicate genetic toxicology methodology to assess nano-safety and show a critical understanding of the development and uses of nano-materials and their impact on human health for risk assessment.

Chapter 1 (Mutations of human health)

This chapter contains

- Introduction to genetic toxicology
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Introduction to genetic toxicology

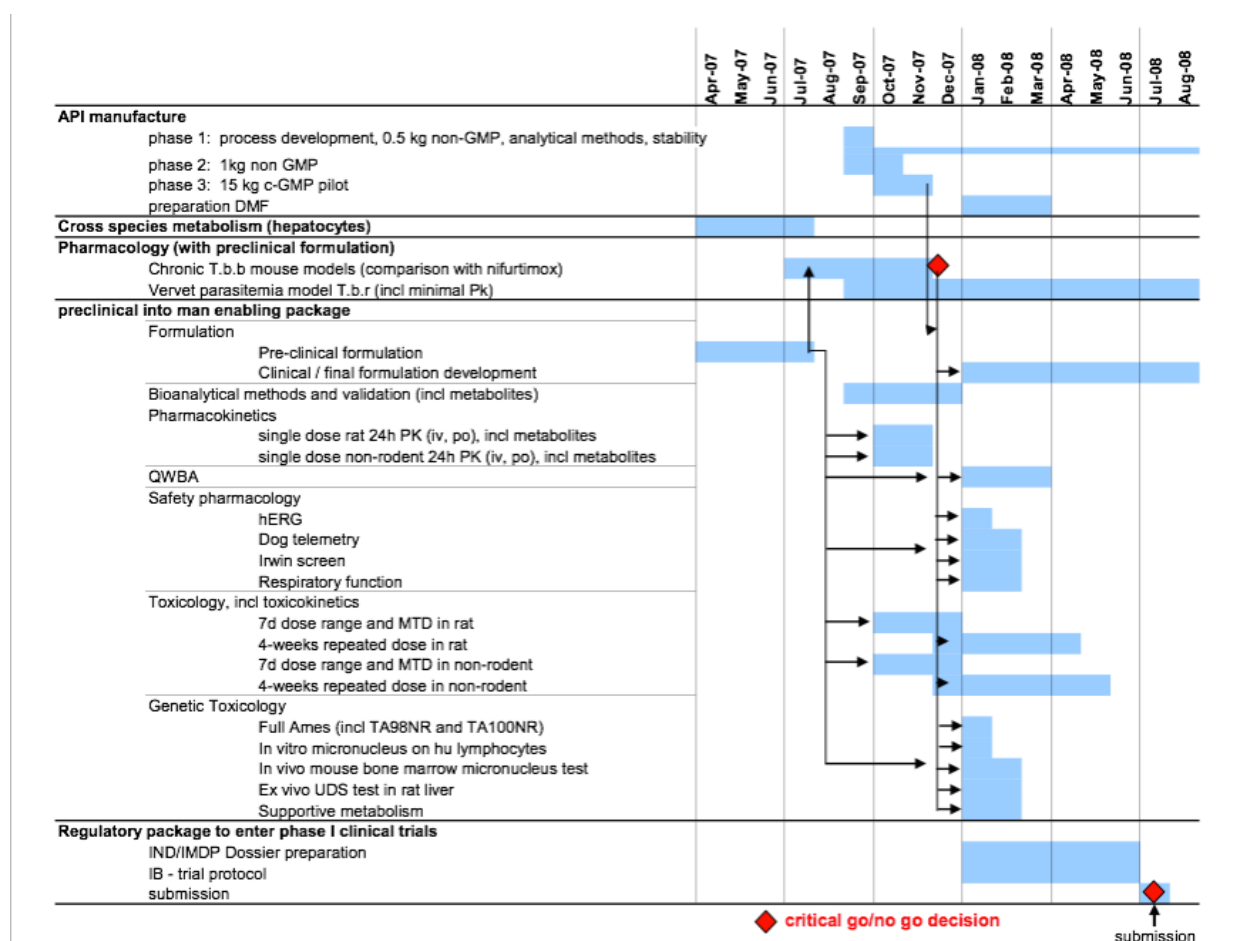
So this topic is about mutations of human health and we are going to be covering these different angles seen below.

- History of Genotoxic agents and how they can lead to cancer. **GEJ**
- Assays with OECD guidelines and potential techniques for the future. **DK**
- Mutations. **GEJ**
- Endogenous and Exogenous DNA damage. **GEJ**
- Nano-safety: nano-materials health impact and risk assessment. **NS**
- Genetic Toxicology in the pharmaceutical industry. **AL**
- Toxicogenomics / Novel treatments for Neglected Diseases. **DT**

Its more of an industry focused moduel and how chemicals that we are exposed to can be assessed and sort of commercialised. We are looking at substances in the pharmaceutical industry and agerochemical industry and we are trying to figure out and prove that there safe and if they are not going to increase our genetic burden then there not going to give us cancer. So to get to that stage are going to have a background of genotoxicity and how it is induced. We then have professor david kurkland coming in talking about the different genetic toxicology assays they use in industry in both in vivo and in vitro and who you do and what you do with the results. It is very useful information and it is used all the time. We will look at some information and how there induced by these substances and how they are produced over time endogenously and exogenously and we are going to look at some examples of how these chemical and how we are exposed to these chemicals both endogenously and exogenously so just inner own cellular metabolism and how we are exposed to loads of nasty stuff as well and why they dont kill us. So nano-safety we will talk a bit about nano toxicology which has quite a big group her ein swansea and we have some of the leading experts here at swansea. Then we will look at some nice case examples to give assume idea of what this type of science is all about and then we will have some working examples for GSK from anthony who will also detail how even if you get a positive of one of these tests. Are you going to bin drug thats going to make a billion pounds a year or are you going to figure a way of showing that its not safe for certain reasons. Then toxicogenomics are quite basic assays that looks at do they cause mutations or not have they got chromosome damage or not and there are some novel ways that toxicogenomics where you can look at gene expression profiles so you get some cells or an animal and treat it with a certain chemical. If you get a chang in gene expression then can you link that to risk as well. So thats for toxicogenomics and lots of people think that thats a good thing and also a bad thing as well. Thats from professor david tweets who is also going to talk about one of his consultancy projects

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where he worked with the drugs and disease incitive, where he looked at some guys worked that got fined in the 70s because it got flagged up in an in vitro microbial gene mutation test as being a mutagen then they went back to and realised it was only bacterial specific mutagen and realised it was wrongg and published papers on it and now its in stage 2 or stage 3 clinicl trials and it has the potential to save about 20,000,000 peoples lives from sleeping sickness and chagas disease as well. So a nice case example about how this paper is actualised in the real life situation so again we have more of an industry focus in this modele and the cancer module is more sort of clincally focused with people form the NHS and such coming in. So heres a nice confusing thing for you that highlights where genetic toxicology comes in. So that is the actual profile from that drug which is actually showing that they carried out an assessment over a time period with this drug called fexinadazol for river blindness thats funded by the gates foundation. We dont care about the drug to much at this stage we just really wanted to see that this is the kind of thing you do. You get a drug, you dont want to carry out one test and then another you want to carry

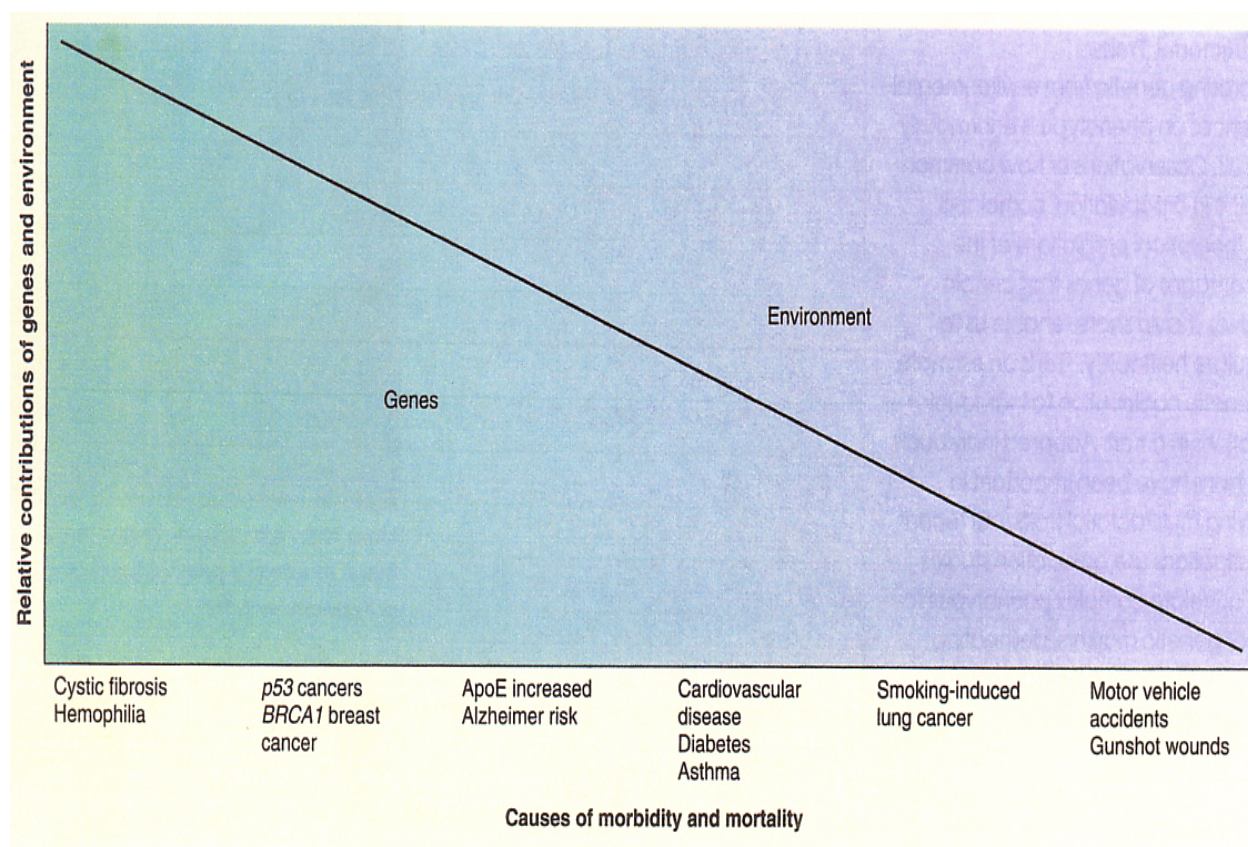


them out all at the same time. So if your going ot market this drug and again its going to be a blockbuster drug giving you around a billion pounds a year. You don't want to stager the safety testing because the longer it takes to get it out the more millions you loose. So the idea is that you do all of these things at the same time and at the end of it all you want is a safety profile to have no flags and showing that everything is noice and safe and that the efficacy shows that it actually works and then you can go on to the stage 1 human trials then. So what we are doing here then is looking at the month by

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month action protocol on how you actually make this stuff in big amounts. The pharmacology side starts going towards the formulation and how to make this stuff to be distributed throughout the system. pharmacogenetics so dose it get distributed around the system quite nicely. Then the safety side is where we come in, in Swansea we dont have an animal house we just do the in vitro side of things. Obviously these tests if its a pharmaceutical substance then you need to carry out in vivo assessment and show that its safe in these animals as well. So here we have things like safety pharmacology, standardd toxicology and these things look to se if these compounds are refining the organs of the animals and then genetic toxicology comes in and just wants to know really are these substances causing gene mutation. Are these substances causing chromsome brakes, are they causing segregation of mitosis causing a loss or gain in the chromosomes. So0 thats what we are doing here and thats just to show us how it lines up with everything else. Then we get all of that we wrap it up together and give it to people like the FDA and they say ok go ahead or they say no and bin the work.

So we are going to look at the history of it all now and look at how genetic toxicology links in to all of this. We will mainly be talking about pharmaceitcal industries but we will elude to some other industries as well like the agrochemical, the petrochemicals and finally the food flavours industry. So what we need to think about is say we we have conditions on the left side of the diagram which is entirely linked to your genetics like the diseases cystic fibrosis and heamophilia. We get over towards this side where the environment starts having more of an impact. This side of things where smoking is quite a nice example. Smoking indues lung cancer where you have some pre disposition to getting increased levels of cancers. You have some strange oncogenes and some strange tumor suppressor genes. But mostly it has been



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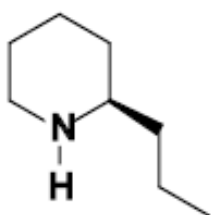
induced by the heavy amounts of smoking and nasty things like polyaromatic hydrocarbons and again we will talk about those later on. So our idea is trying to figure out how the environment can impact on the genes really. So why did all of this start out, it started out with an interest in the underlying mechanisms of mutagenesis and realising that radiation and chemicals are actually doing some strange things. You get some cells and start playing around with them and you realise that radiation and chemicals start killing the cells and starting to make them divide in strange ways. Then later on in the world wars it was the era of the atomic bombs that had some issues that were causing people to get cancer and they started thinking about what the mechanisms of this could be. There is mustard gas as well where they saw it as a blistering agent in the world wars. All these blistering agents do is enter the body and attach to parts of your DNA and then inhibit the cell from dividing. So those cells die and then they lyse (necrosis) and this causes a blister. These sorts of things have some side effects as well. So because people wanted to get an understanding of this they started getting a bit worried as they are being exposed to many more agents than was previously thought which is when regulations were starting to get introduced. So the identification of the role of these mutations in cancer induction or progression. So people started understanding that these nasty chemicals and substances caused cell death and cancer and then people



started getting appreciation of why people were dying from cancer. So a bloke seen here below is Philippus Aureolus Theophrastus Bombastus von Hohenheim said that everything is poison and there is nothing that we know of that is without poison, it is only the dose that can commit something to be considered not poisonous. This comes up quite a bit and even recently where you hear in the press where you hear every week saying that something gives you cancer and then the next it doesn't give you cancer. This is really all about the concentration and all about the system people are testing it in. So even nowadays people are still arguing about this sort of thing where they realise that if someone is exposed to a nasty chemical like a bottle of coke then there are loads of compounds that make up the coke and the bottle that if we broke down we would find are carcinogenic but because there are at such low levels they don't give

us an effect. So you can scare people but really you have to have some realistic understanding to realise that the concentration is really the important factor. This is not so much genetic toxicology and is more toxicology and even before Christ so BC people were using

chemicals to kill each other as depicted in the diagram below which is the killing of Socrates by Hemlock.



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this was done with an alkaloid called coniine which could cause paralysis, convulsions and eventually death. So this would lock open the synaptic gates cause a paralysis and that was that. So even back in these old days people had an understanding of what the substances were and people also had an understanding of what cancers were, which we will see in a minute. So even the dinosaurs were said to have types of cancers where their bones show evidence of cancers. The Egyptian mummies were shown to have one cancer also. So you see that cancer is prevalent throughout history and the only thing that's prevalent about the Egyptians is that they started getting a bit of an understanding of some types of cancers. So in Egypt there is evidence for 8 different breast tumours and ulcers and they actually came out of a depiction that was drawn on a papyrus. So they came in with a looped bit of metal and they would cut off a bit of tissue. They would see that there is some sort of tumour in these breasts, heated up a bit of metal and chopped it out and treated cancer even before in these early days. So there has always been this big understanding of cancer and the understanding that if you chop it out then that's a good thing.

So the word cancer comes from Hippocrates who named these tumours as carcinomas or as carcinos because they resembled a crab with these spreading out regions. We learn more about that pattern in a different module but really it's due to over-vascularisation or these tumours. So back in 1500s people started chopping each other up and seeing a better understanding about cancers and then with the invention of the microscopes they saw at the cellular level the changes in these types of cells. So they started doing pathology and seeing that in advanced cancers the cells are completely changing their morphology from one cell type to another. These are the sort of advancements you need these things like microscopy. Later on they started linking occupational exposures to certain types of cancers as well, so we got occupational cancers showing that a high incidence of breast cancer was seen among nuns. So because nuns weren't actually sexually active and didn't have children then they didn't have these periods in their lives where their menstrual cycles were stopped and they didn't have these changes in the hormone levels. That's been linked to the increased levels of breast cancer in the nun population.

There are also nice papers on this in the Lancet. Later on all the substances that Swansea is more associated with are Percival Potts showed that in a certain type of industry people were getting certain types of cancers were these chimney sweeps. So they were exposed to these high levels of poly aromatic hydrocarbons. If you burn something like a cigarette or coal or whatever then you get these poly aromatic hydrocarbons. These people up a chimney all day would ingest this stuff and they were getting increased levels of this type of cancer. The Percival Potts being the earliest one characterised. Later on people started getting



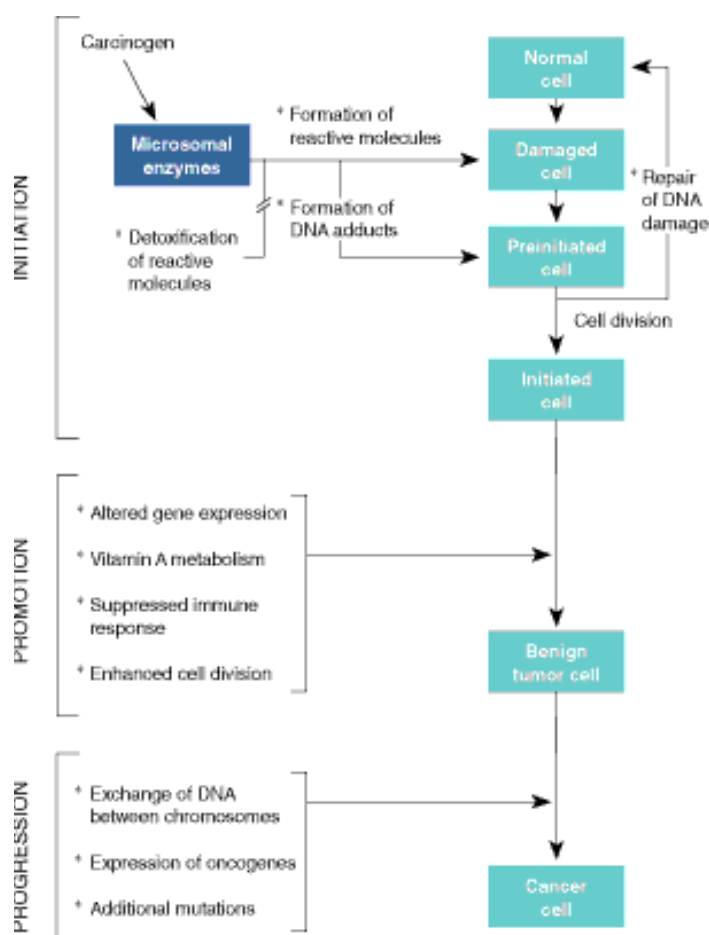
bit more obsessed with fancy dyes and these azo-dyes which are things that are used in cloths and used in lots of products to make them colourful. These Azo-dyes were actually giving cancer as well. So lots of types of cancer but these are a few examples. They started linking in these working industries to getting increased levels of cancer because of their increased exposure to Azo-dyes. So a lot of the time again in the press you may hear about increased risks and things like this and it's not normally the actual

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population that's at risk is the people in the work houses that are being exposed to the high levels that should really be worried about the whole thing. So later on we started seeing these things like coal tar, people used to treat exa with coal tar and this has got insanely nasty stuff in and if your rubbing these things on to your skin then you are getting these poly aromatic hydrocarbons in your skin. Treating animals with this and rubbing it on animals actually gives them skin cancers. Then they started getting so interested in it they actually synthesised Benzo-a-pyrene which is a super carcinogens that is in cigarette smoke and then when you synthesis it your not testing the big mixture of everything your testing just that one chemical you can get a really good idea of whats going on when you can synthesis it and get a really good idea about whats going on when you can synthesis it and test it specifically. Then the whole field started to develop and they started realising that if you got model animals and started treatng them with nasty stuff then you could actually induce things like cancer. So we are not going to go into a lot of depth about what cancer is but it is useful for this lecture.

So its the uncontrolled multiplication of cells so what we are thinking about is chemicals which lead to this uncontrolled multiplication, lead to mutations in proto-oncogens and tumor suppressor genes which are going to make your cells divided in a strange way and the terms we use are things like benign and where a cancerous cell is contained in one place and malignant when it gets worse and they can spread to other areas and cause things like metastasis. So cancer in humans, the reason why we are going to be talking about these different terms is because you can have substances which are cancer initiators and cause and initiation of cancer. So these are like the first mutation in a certain gene, promotion where theres an accumulation of these diffenret mutations in the different genes. Progression where there the cells are actually changing into different types of cells and malignancies when they start to migrate. So how we asses this in in vitro and in vivo well we can use cell lines, transform cells and the easiest way of seeing this is dose this substance cause a tumor in the experimental animal and what we are talking about in

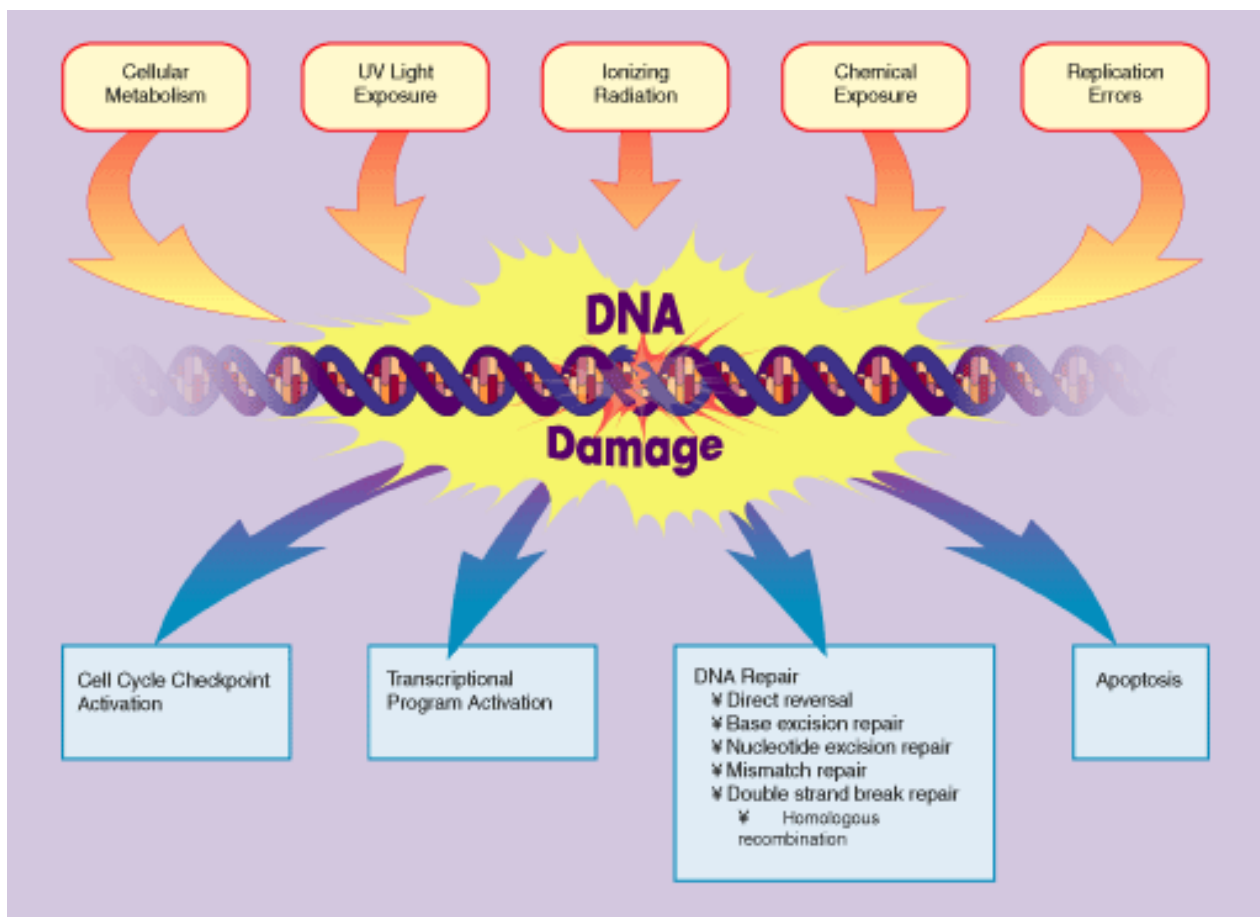
this module is other assays so you don't have to carry out these assessments in animals. which is good because the bioassay requires 400-800 animals and takes two years to compleat. Genetic toxicology takes no animals and much less time so these are just some of the benifits. So we will be talking about some of these things. So what causes cancer basically every thing causes cancer. Organic chemicals things like asbestos, there are hormones as well like 17beta estradiol so even our own hormones which we produce endogenously can cause us cancer as well. With hormones there are lots of substances that



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can mimick these hormones which we said can have deleterious effects. Then mixtures, which is quite hard to test but then mixtures of things can give you cancer. too.

So hormones are quite a big groups that we will be going back to throughout this module some inherited conditions and really we will be talking a lot about reactive oxygen species. So these are things like the hydroxyl radical and these can be induced by many different things like increased stress and lots of irritation. Lots of the antioxidants try to sell you are oxygen scavengers which can just come along and soak up these reactive oxygen species. One of the main deleterious effects are from ionising radiation and it causes reactive oxygen species in close proximity to your DNA. So there's water next to your DNA and ionising radiation cause these to turn into the hydroxyl radicals and this causes you to get some DNA damage but again we will get some information about this later on. We have also got some external factors as well so we have our habits like smoking and the diet, so your own lifestyle can mean your exposed to higher levels of these certain types of chemicals. Viruses as well can come in and can cause DNA damage and get in between the bases in your genes. chemicals and radiation. So basically we are exposed to a bunch of different things and were not all dead. So there is a lot of preventative measures to. So if we go along the top of the diagram here things like cellular metabolism where just in the mitochondria we metabolise chemicals and when you metabolise something your likely to get some sort of reactive byproducts. UV light exposure is linked to cancers things like cyclobutane pyrimidine dimers they come in and make two bases join together just through photo reactivation. Ionising radiation we will talk about a bit, even in this lecture but mainly chemical exposure because we are talking about the



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pharmaceutical industry they are making chemicals and they want to make sure that its safe. So thats what we will talk about quite a lot. The replication errors so when the DNA is trying to duplicate itself, you can actually mess this up and cause some mage just endogenously. But as we said we are not all dead we haven't all got cancer because there are some nice mechanisms to make sure we don't get cancer. So things like cell cycle checkpoint activation, if you have got some DNA damage it gets picked up in the cell cycle checkpoinnt, it stops at the cell cycle checkpoint and give the cell some time to repair that damage or giving the cell time to induce apoptosis which is the programd cell death. So in this case if we are talking about cancer in this case it is a good thing because we are not going to produce a daughterr cell with the increased damage,,So cell death to cancer is actually a good thing because it is preventing it. Then shen you have this damage you can activate some different programs as well. The up regulation of certain genes and pathways with DNA repair being one of the main ones. If you have a substance like a reactive oxygen species like a bento-a-pyrene di epoxide interacting with the DNA. If its pickd up then we have some very niceness and tidy repair mechanisms that have evolved naturally that can get that DNA adduct and remove it in a clever way. So we will look t some examples now and also some temrinology. So these terms will be used throughout the module to keep note of them really:

Genotoxicity = these are really the studies of the chemicals that causes toxicity to your genes. That is the all encompassing term for everything else written below.

Mutagenicity = More specifically the things things that cause a gene mutation or some kind of base pari change or causes a deletion or just one base pair to become another base is mutagenicity caused by mutagens.

Clastogenicty = This disruption of a chromosome changes or some thing that causes chromosome brakes is called clastogenicty.

Aneuploidy = being linked to down syndrome is basically a change in the number of chromosomes so something that can induce this would be called an anuegen causing aneuploidy. So this is education of numerical chromosome changes such as the gain or loss ozone chromasome or the gain or loss of multiple chromasomes.

So one of the early mistakes and for which products are so heavily regulated nowadays that were made were the spreading of DDT or dichloro-Diphenyl-Trichloroethan. This wasted as a pesticide to try and fight the vectors of malaria so it actually killed off insects and particular in these areas of africa which carried these different types of disease. A pesticide for vectors of malaria, typhus, yellow fever and sleeping sickness.DDT was really quite scuffle in killing of these insects in quite a good way. the problem is that it had some side effects particularly when tested in vitro and in vivo where it was seen to be a probable human carcinogen and seen to be an endcrine disrupter as well. So it actually mimicked hormones it was affecting the estradiol receptor and the androgen receptor. So this chemical that they were using to kill off all these insects were actually coming in and interacting with these receptors and the thing is here that these are hormone receptors and they are also transcription factors and they regulate

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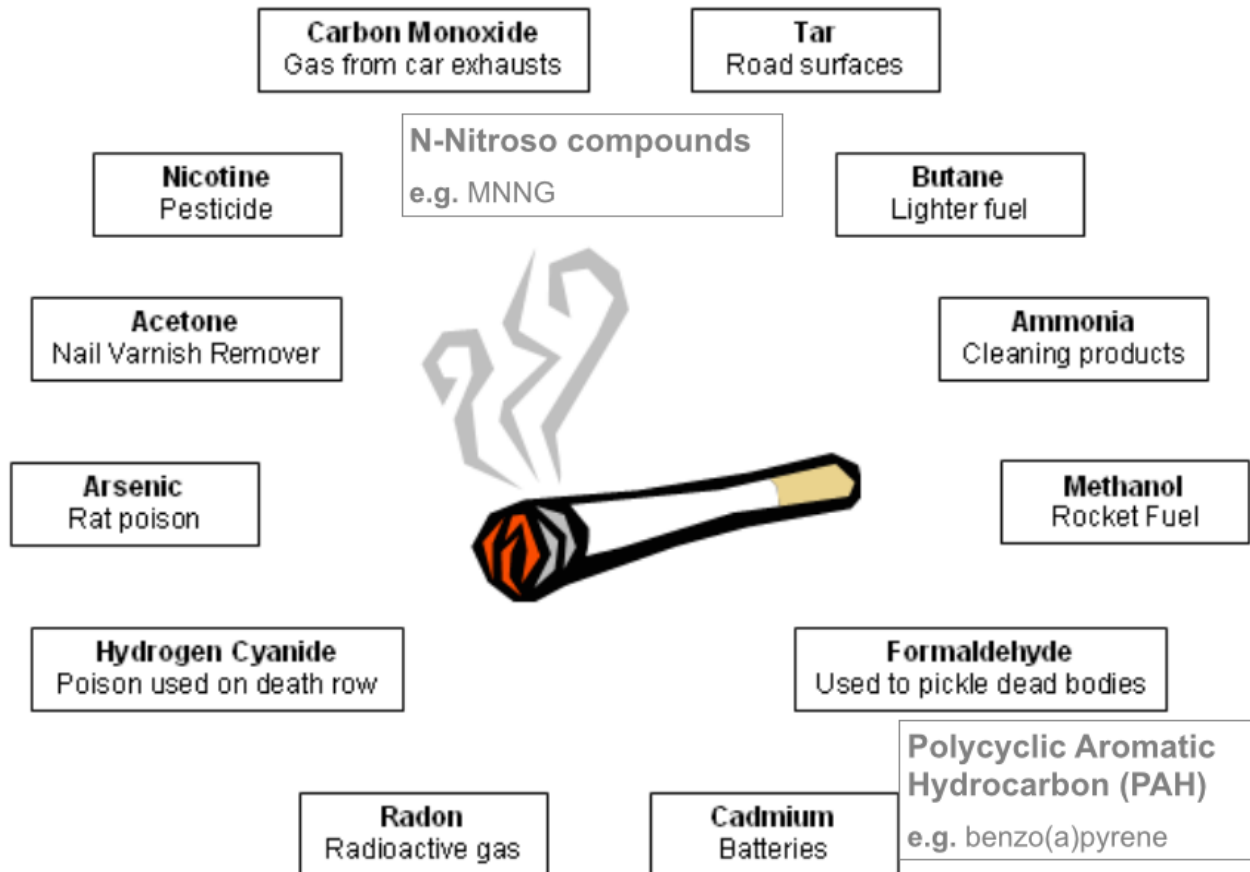
absolutely loads of genes. So if you start messing around with these then you are going to have loads of effects. So this was back in the seventies, these were really quite massive treatment programs were people were being exposed to quite high levels of DDT. Then they started testing this quite heavily and



This machine is spreading a kind of fog of DDT spray to see if it will kill the mosquitoes and other insects on the beach. Outdoors, the spray soon thins out and does not harm people.

saw that the animals systems cause liver cell tumors and its causing liver carcinomas and hepatic sarcomas and lung carcinomas. So there is really a conflict nowadays where they are saying that they didn't see a huge increase in human cancers in these areas and this is probably due to the levels or the expression levels. We wont go into to much detail there we will just realies that this is one of the main reasons that chemicals and compounds are so heavily regulated at the moment. Because you get problems like this and realise that everyone is being exposed to these nasty chemicals and this si the type of ting thats happening and even beaches in clifornia. where they would go up an down the beaches and just spray this in people faces. This made every oen happy because althoe flies are dining but when you see that it causes liver cancer and it messes up your endocrine pathways its not really something that you want to be spraying yourself or your kids with. Something else we quite happilly expose ourselves too a lot of the time are cigarettes and the byproducts of burning things like these N-Nitroso compounds and all these poly aromatic hydrocarbons seen below. We have got things like bento-a-pyrene which we keep going back too. Obviously there are al of these other things but the main one we will be looking at is the main one thats been associated with the lung cancers and in the cigarette smoke are these poly aromatic hydrocarbons. So because of this we have got a quite good understanding of human

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experiments reall. In do these chemicals cause cancer, yes they do and we can test these peoples tissues for deleting the levels in these tissues we can actually detect the types of DNA adducts that are being caused and we can see someones lungs cancer then we can see specific elements or adducts produced by the substance in the lung cancer and we can see that this substance is inducing lung cancer. Theres auto mutation profiles as well so a very clever way linking that types of substance to those increases in cancer a nice thing here is when we mix our drinks in with coffee, a lot of these compounds are in coffee as well but again a very low concentrations. On the levels within coffee as well we have bento-a-pyrene, hydrogen peroxide, formaldehyde, benzene. This is in coffee which is a product that we drink quite a lot. If this was a new product then it would be band strait away but because of its industry and history then it cant really be banned. So we realise that we are exposed to all these different things on purpose and the also because we don't really know what were ingesting as well. So because of that we are exposed to these different things its mostly environmental factors in cancer deaths, diet is the biggest cause of cancer and especially because of the increased levels of estradiol because estradiol is made from the afatty substance that are being increasingly ingested. So with an unhealthy diet you get

APPENDIX TABLE III

Carcinogenicity in Rodents of Natural Chemicals in Roasted Coffee⁸⁸

Positive:	
N=19	acetaldehyde, benzaldehyde, benzene, benzofuran, benzo(a)pyrene, caffeic acid, catechol, 1,2,5,6-dibenzanthracene, ethanol, ethylbenzene, formaldehyde, furan, furfural, hydrogen peroxide, hydroquinone, limonene, styrene, toluene, xylene
Not positive:	
N=8	acrolein, biphenyl, choline, eugenol, nicotinamide, nicotinic acid, phenol, piperidine
Uncertain:	caffeine
Yet to test:	~ 1000 chemicals

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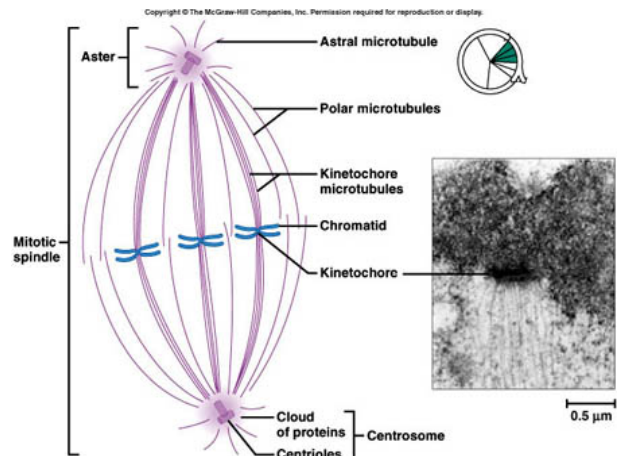
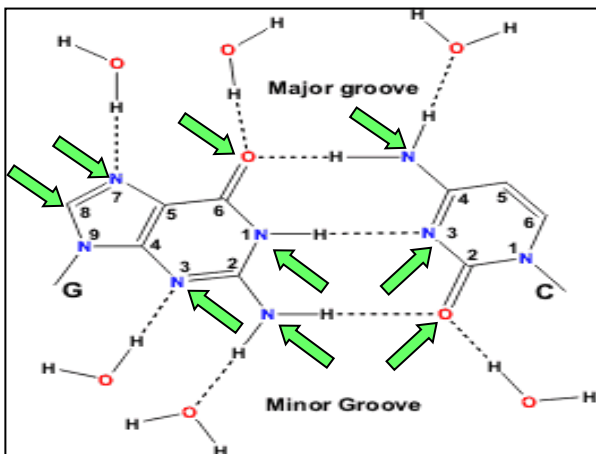
increased levels of cancer. Tobacco is responsible for 30% of these environmentally induced cancers. Infection as well if you have repeat infection then you actually get a lot of things like reactive oxygen species being released in that area and you get different genes unregulated. So infection and persistent infections can cause cancers as well. There is a table below where you can also see the less prevalent cases of cancer below, so mainly our diet but there are also other things as well.

Diet	35% (10-70%)
Tobacco	30% (25-40%)
Infection	10% (?)
Alcohol	3% (2-4%)
Reproductive and sexual behavior	7% (1-13%)
Occupation	4% (2-8%)
Pollution	2% (<1-5%)
Geophysical factors	3% (2-4%)
Medicines and medical procedures	1% (0.5-3%)
Industrial Products	1% (<1-2%)

Looking at the bottom you can see that medicines and medical procedures is responsible for around 1%.

So the mechanisms of action and their relation to cancer. So we are looking at chemicals that cause genetic damage and the relationship with that genetic damage with the cancer. This involves three classes known as the

1. aneuploids = cause changes in chromosome number, either gains or losses.
2. Clastogens = Some kind of chromosome break
3. Mutagens = Some kind of mutation



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You can get different classes of chemicals as well that directly interact with your DNA. All the interaction with things like the cellular machinery with DNA replication machinery. So things like Benzo-A-pyrene and the restive oxygen species can come in and attache themselves to the DNA. The other ones like the anuegens come in and attach to things like the mitotic spindles here. So if you get a chemical that comes in here messes up the mitotic spinal, messes up the other one meaning you are going to loose that chromosome. If you mess up both sides then both sister chromatids will go to one side. So you can see how the chemicals can come in and cause these genetic effects. So these anuegens are the reason for the induction numerical chromosomes changes, we are going to learn soothing a bit confusing as well now. So lots of these chemicals here we are going to look at how they give you cancer but lots of these chemicals are linked to chemotherapy as well so its all about concentrations. So the chemicals that we will look at have low concentrations that don't do anything, high concentrations kill off all the cells, concentrations in the middle can cause cancer. So its all about concentration. So the induction of neumerical chromosome changes. So these anuegens are things that affect the cell division and the cell cycle apparatus. To get these gains or losses of genetic material, this is a hallmark of cancer where you could get the gain of a whole extra chromosome and if there is something like an oncogene on that chromosome then it means that oncogene is going to be overly expressed on that chromosome because you will have three copies not two. When it gets hevily expressed you get things like polyploidy, so instead of getting two sets of chromosomes you get three sets of every chromosome. This is going to have the same sort of effect where you are going to get massive up regulation of loads different genes and many of them associated with cancer. So these engines potentially lead to cancer, if your germ cells are exposed to these engines then potentially it could lead to things like down syndrome. So if the sperm or the egg are exposed to things like these anuegens then you get these non-disjunction type of event occurring and potentially that can lead to down syndrome and in the offspring. So the mechanisms tic action of these spinal poisons disrupt the depolymerisation and disrupt the spinal polymerisation or they can mess around with the distribution

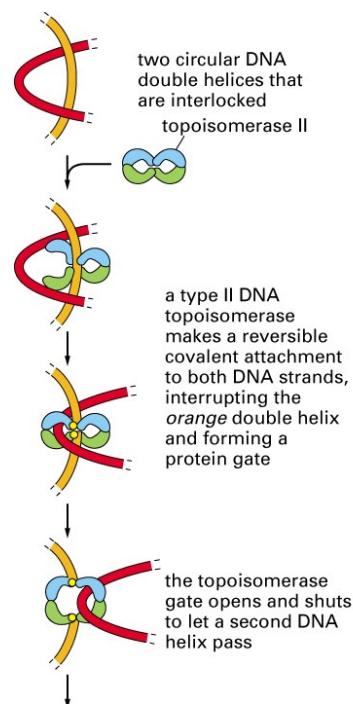
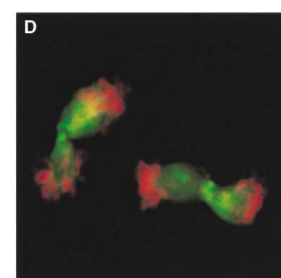
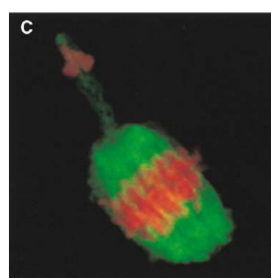
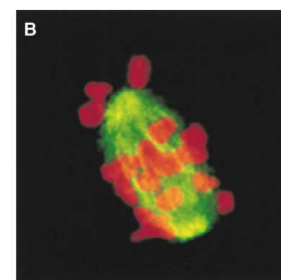
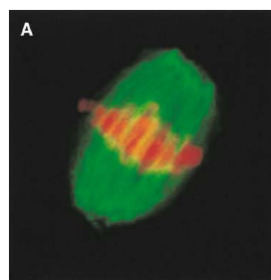
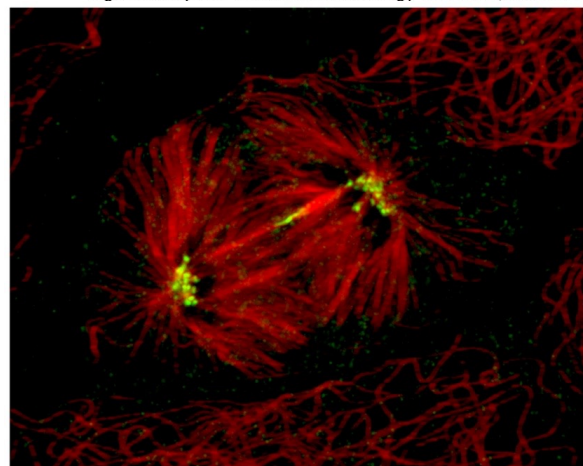


Figure 5-27 part 1 of 2. Molecular Biology of the Cell, 4th Edition.



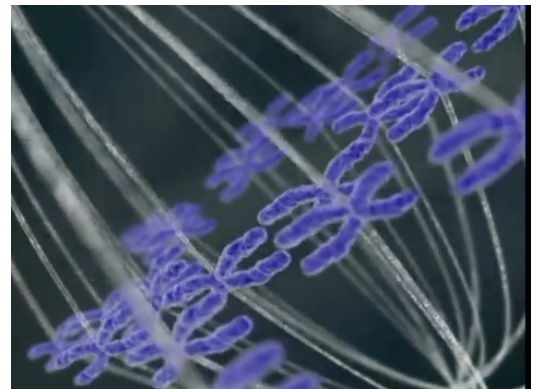
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of these spindles as well. So you can get cells to divide into two like this by getting them to pull the spindles to two different sides as seen in the diagram above. You have three or four of these centrosomes that can pull all the DNA into three cells, four cells instead of two and things like biphenyl A which is present in coca cool can do that. So in the video we are looking at the we are talking about chemicals that can mess up the diffenret processes.

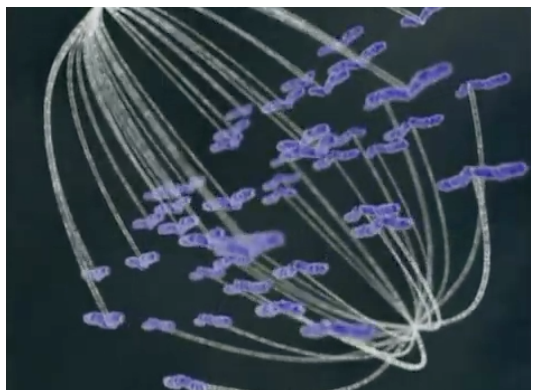
We are looking first at the mitotic spindles coming out from the poles and a chemical can actually stop this polymerisation form happening.



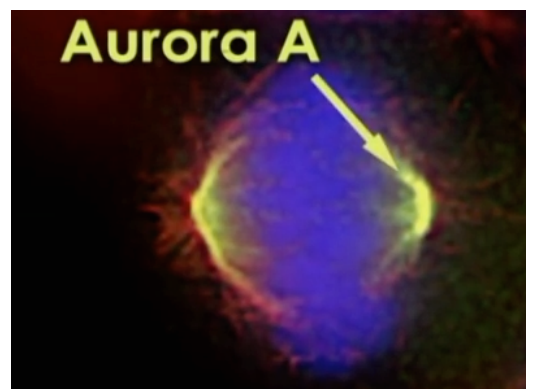
Its a complex process were for every cell division this needs to work properly. The mitotic spindles need to migrate and correctly bind to the sister chromatids at the kinetochores



We need to separate the sister chromatids to opposite end of the cell and there are chemicals that can mess these things up.



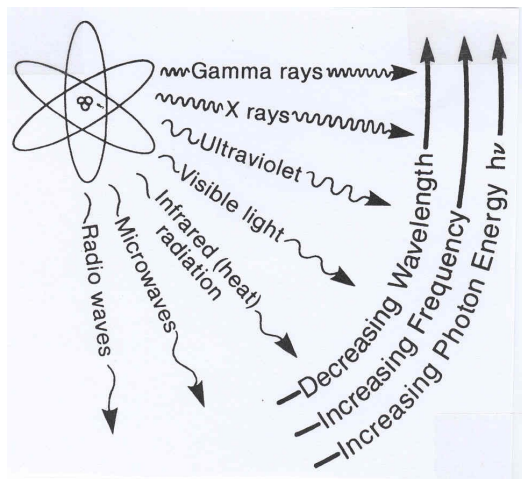
So if you mess up one of these poles then they you can get defects in the Auroras and you can get incorrect division of these chromatids. So just by exposing you chromatids to these anuegens will increase your chances of loosing your chromosomes into the cytoplasm and loosing chromosomes from the cell to your daughter cells. So not really something you want to be exposed too and really



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something you don't want your germ cells to be exposed to either because it can cause these cells that are reminiscent of down syndrome and things like that. The kind of places that these things appear. You have a medicine Colchicine used to treat Rheumatic diseases and especially gout which is a highly deadly aneugen. There are some things in bread that can get metabolised in to these engines that are quite happily ingested but again concentration is everthing. So the concentrations we are ingesting makes it ok. Bisphenol-A which has just been reviewed by the europeans increased aneuegency has just done the risk assessment on bisphenol-A and said that the levels we are actually exposed to is safe. So it was banned in baby bottles a few years ago. This is because thereby bottles were being but into the microwave and then all the bisphenol-A form the plastic would lack into the babies milk and then the babies were ingest loads of it. So shown that we are normally exposed to quite low concentrations of it. So its all around the place but in low concentrations is the message really. When your assessing this you can see that bisphenol-A dose do these things and it messes up the chromasomes and you get chromosome loss here. It mimics the hormone estradiol in the body causing things like disruption in the hormone pathways. It induces things like chromosome lagging and the chromosome saint attached to the mitotic spindles. Because they mess aroundd with the cell systole so much and they mess around with cell division so much they are quite good chemotherapy drugs at high leves. So at higher levels the cells are trying to divid into two and at a higher level then the microtubules are actually damaged the cels cant divide into tow. So if the cel cant divide into two then your not going to get a big ball of cells forming a cancer because they can't divide. Also if they are trying to divide and they cant a lot of cells just kill themselves so lots of these problems occur then this si a good thing. This is also a bad thing because there not cancer specific. So they kill all your dividing cells in your body which is why you get things like hair loss and internal bleeding from your veins being less stable. The plus side though is that it dose kill the dividing cells in you tumours. So what it doe is induce mitotic arrest in the cells are just stuck in mitosis and this is when you want apoptosis and cell death to occur so you want the tumour to die due to this inability to divide. apoptosis in interphase can also occur as well. So although these anti mitotic processes are becoming affected in cancer chemotherapy they use this limit on molecular effects on tumours of the development of resistance. So sometimes there resistant and the middl of the tumor isn't being exposed to these chemicals and people don't like the dividing effects of all the other dividing cells dining off as well. So there a bit too much of a blanket approach but if you use them in some sort of targeted therapy where people in nano-medicine will say that you can targets drug using some kind of magnets or immunoglobulin ways to target a molecule to your tumor. It brings in high levels of a nasty drug like this so you don't get all of your other cels in your body exposed to this and just the specific tumor ones. So these drugs are used for loads things like the beta alkaloids and the taxes, things form plants so there natural.

So clastogens ar ethe ones that cause chromosome brakes and again a los of genetic material through this chromosome brake is best characterised by ionising radiation such as gamma, X-rays and certain types of UV. Theres some good data on this from the human population so we have one of the best examples is when they first found out the radiation is causing cancer. So in the old days with the old fashioned watches they used aradioactive pants so that your watch is flouresent in the dark and normally the ladies that were painting this on to make a nice clear paint brush when your painting its quite nice to



	non-ionizing			ionizing	
Type of Radiation	extremely low frequency	radio microwave	infrared visible light	ultraviolet x-ray gamma rays	
Effects	non-thermal induces low currents ???	thermal induces high currents heating	optical excites electrons photo-chemical effects	broken bonds damages DNA	
Source	static field power line	AM radio FM radio TV oven	heat lamp tanning booth	medical x-rays	

lick

the end because it means that you can do some nice fine painting. They were licking a radioactive paintbrush and getting levels of young cancers. So it was really linking the radioactive paints to these young cancers. So types of levels that we are exposed to mainly from radon and if you live in the west country where there are high levels of granite then you're going to be exposed to higher levels of radon and when you're exposed to these sorts of levels of radon and these sorts of levels of cosmic rays then the people who work in the medical field are exposed to higher levels of X-rays and also in radiotherapy as well. So we are exposed to low levels of radiation but we are not all getting cancer from radiation. Some people who were getting cancer from radiation in this example. You can't really do human experiments where you expose people to radiation and see what happens so the closest control experiments that we have on human experiments are things like the nuclear reactor disasters like Chernobyl. So what happened in 1986 was Chernobyl had an unauthorized experiment where they were playing around with the reactor. It got too out of control and it exploded and released lots of this radioactive material onto the surrounding population. So around Chernobyl there is quite a strong south westerly wind so the cities of Ukrainian-Belarusian were exposed and then a few days later it went easterly and came over towards the UK as well but that was after a couple of days when there were lower levels. So these people were exposed to quite high levels of radiation compared to the background levels. Particularly the liquidators who were the people who went in to clean up the mess straight away. They got exposed to the highest levels of radiation. Then the evacuees were the people who had to escape somewhere safer. The next ones down were the residents of the surrounding countries. So people were exposed to quite high levels of radiation so everyone thought these people were going to get cancer and their offspring would get cancer and have congenital abnormalities. The main issue really was thyroid cancer. This thyroid cancer affected mainly children in adolescents because the radioactive iodine is released here and the radioactive iodine released from the reactor was eaten by cows,

Figure 31. Radiation Hotspots Resulting From the Chernobyl Nuclear Power Plant Accident



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Population (years exposed)	number	Average total in 20yrs (mSv) ¹
Liquidators (1986–1987) (high exposed)	240 000	>100
Evacuees (1986)	116 000	>33
Residents SCZs (>555 kBq/m ²)(1986–2005)	270 000	>50
Residents low contam. (37 kBq/m ²) (1986–2005)	5 000 000	10–20
Natural background	2.4 mSv/year (typical range 1-10, max >20)	48
Approximate typical doses from medical x-ray exposures per procedure:		
whole body CT scan	12 mSv	
mammogram	0.13 mSv	
chest x-ray	0.08 mSv	
[1] These doses are additional to those from natural background radiation.		

concentrated in the milk and drunk by the children and because they were in these deprived areas they already had iodine deficiencies. So soaking up massive amounts of iodine all of the radioactive iodine into their thyroid glands, this caused people to have mutations that lead to increased thyroid cancer in these individuals. Really when the WHO looked at this they thought there weren't as high levels of cancer as they thought and there were not nearly as many as predicted levels of congenital abnormalities as you would have thought in the offspring of people who were exposed. Really they are convincingly high number in the general opinion where there're a lot of increased levels of cancer. So the model to predict that by 2065 about 16,000 people will have thyroid cancer as a result of this disaster and because there is only about a 0.01% of all cancer deaths this is deemed to be quite a low number. The 25% of other cases that are expected to be due to radiation from this accident but because people say this is a low number then it's not that significant. Really people do have a case here because there were so many people exposed to the radiation and you would expect much higher levels of cancer. But the WHO says that it's not as bad as they previously thought, and some people say it's a good thing because there are loads of animals in this area now and they survive just fine. So there are loads of molecular mechanisms of these clastogens there breaks and rearrangements cause these genetic instabilities and they can lead to increased mutations. So the thing about clastogens is that when something is classed as a clastogen it's all a mutagen as well. So normally it's a clastogen and the substance is so reactive it reacts with the DNA causes some kind of strand break. When something reacts with DNA is also likely to cause mutations as well. So a lot of the clastogens are also mutagens. We are exposed to loads of these things as well, so occupationally chemotherapy nurses and doctors are exposed to these things. Obviously the chemotherapy patients will be as well. When we're rethinking about why people get cancer and how they get exposed to these carcinogens, there's a risk-benefit calculation where if someone is going to die within one year of cancer if they take this drug then they'll die in ten years from taking the drug. Then the risk

- Short-lived ^{131}I (Iodine),
 - Long-lived ^{134}Cs (caesium) and ^{137}Cs
- Models predict that by **2065 about 16,000** (95% UI 3,400-72,000) cases of **thyroid cancer — 0.01%** of all cancer deaths.
 - And **25,000** (95% UI 11,000-59,000) cases of **other cancers** may be expected **due to radiation** from the **accident — 0.015%** of all cancer deaths.

Nature (2006). 2006. **Special Report: Counting the dead**

benefit of them taking the drug outweighs the risk. So there is a risk benefit thing that comes into play when you talk about a drug and particularly for one of these terminal illnesses. These sorts of risk benefits don't come into play when talking about agrochemicals like pesticides or insecticides because so one wants to die in ten years time because they have eaten the cabbage discovered in these things. So there is a risk benefit factor that only really applies to medicines. We are exposed to these different things, these mutagens, occupational through people smoking. Again environmental exposure and again food hopping up all over the place. Our bodies actually induce quite high levels of these things as well and one of the colleagues in amberca where if you do the risk assessment of breathing the environmental protection agency would actually ban breathing because there are high levels of chemicals that you shouldn't be exposed to. So they can get a bit over the top with these calculations. These insecticides we do get exposed to. So the kind of chemicals that cause these mutations are something becoming and usually drop a bit of themselves onto the DNA and this case it's an ethylating agent which drops an ethyl group onto the DNA. Upon the DNA replication this is misrecognised and the chemical that causes this DNA adduct to occur is misrecognised so upon the replication you get a strand base pair change. So some other things that cause this are things like cisplatin so again the chemotherapy drug regularly induces mutations, is believed to cure cancer by binding to the DNA and interfering with its DNA repair or its repair mechanisms. It binds to two places in your DNA. Cisplatin binds to two places in your DNA and causes mutations as well. So when we are thinking about genetic toxicology we're talking about chemicals that cause gene mutation and chromosome breaks and chromosome loss. You need to be aware that there are some others as well like non-genotoxic carcinogens that don't cause these gene mutations but things like these anti-estrogen ones. So they don't cause DNA damage but they can mess around with your hormone pathways that can cause uncontrolled cell death as well but again risk benefit means that this can be a very helpful drug treating many different types of cancers.

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So there is a bit of a paradox with these hormone drugs like tamoxifen. So it is actually genotoxic and it is a carcinogen but it is not a genitor carcinogen because the DNA adducts that it uses don't cause the cancer. So this is a bit confusing that it flags up that it doesn't react with DNA and it is also a carcinogen then you would immediately like those things together but in this case where this is not actually the case by causing cancer through other mechanisms. So it's a bit of a weird one and leads to arguments.

So those are things that react with the DNA and potentially react with the DNA or the cell cycle machinery. Another type of chemicals that are important to look at not so much in the genetic toxicology pathway but in the reproductive science pathway will be teratogens so usually these things mess up some sort of developing pathway in the developing foetus. They would so much mess with the DNA but rather something at the protein level and so teratogens like Thalidomide where the pregnant mother will take these substances and it will cause some malformations in the development of the foetus. So Thalidomide was the best example of this where it was given to pregnant mothers as a morning sickness drug and then they realised the offspring of these people who had high levels of this treatment actually had this stunted limb growth. So limb bud development was affected. So what it does is it doesn't react with the DNA it reacts with cereblon important for this limb formation (it actually reacts with the Shh pathway and the reduction in cholesterol). So there are loads of other ones of these as well so when you're pregnant and someone says don't take this it is mainly because it can act as a teratogen. So the same is true for all of these things in the table below:

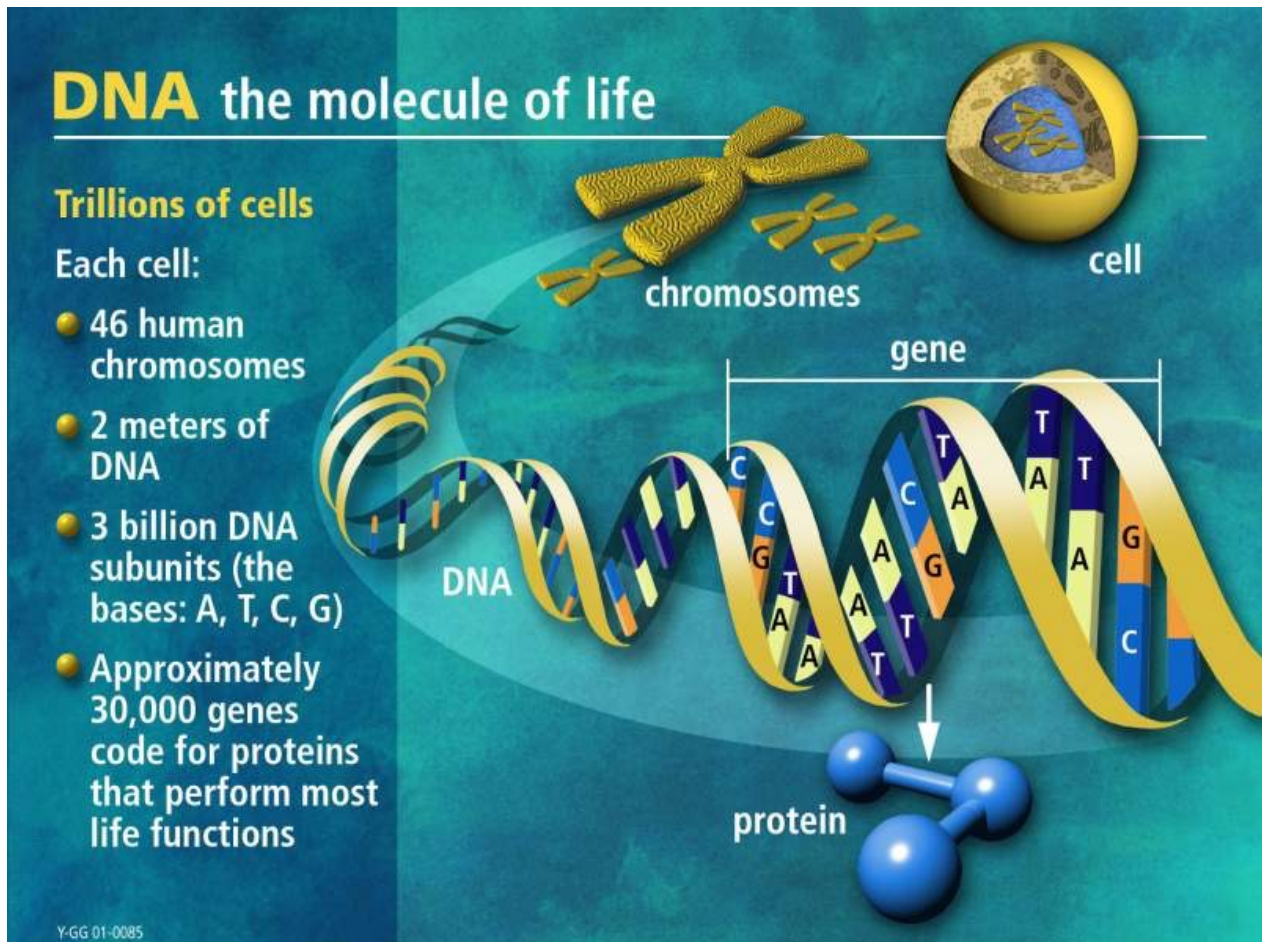
Type	Class	Agent	Risks to Embryo or Fetus
Chemicals			
Pollutants	Heavy metals	Methylmercury	Teratogen
Recreational drugs		Ethanol (alcohol)	Teratogen, growth retardation
		Nicotine (tobacco)	Death, growth retardation
		"Crack" cocaine	Death, growth retardation
Over-the-counter drugs	Salicylate	Aspirin	Death, growth retardation
Drugs			
Anticancer agents	Folic acid antimetabolites	Methotrexate , Aminopterin	Teratogen
	Alkylating agents	Cyclophosphamide , Busulfan	Teratogen
Antibiotics	Tetracyclines	Tetracycline	Hypoplasia and staining of enamel
	Aminoglycosides	Streptomycin, Kanamycin	Hearing defects
Anticonvulsants	Oxazolidinediones	Trimethadione, Paramethadione	Teratogen, death, growth retardation
	Hydantoins	Diphenylhydantoin (phenytoin)	Teratogen
	Carboxylic acids	Valproic acid	Teratogen
	Bromides	Bromides	Growth retardation

Chapter 2 (Genetic in genotoxicology)

This chapter contains

- Introduction to genetic toxicology
- Ames tests

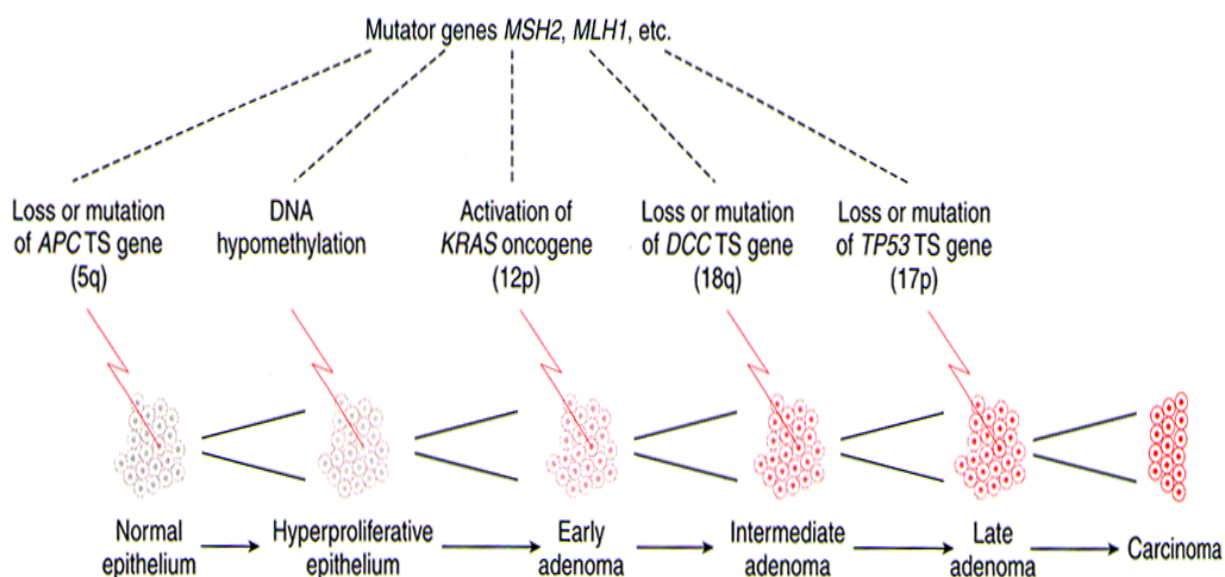
Introduction to genetic toxicology



So we are talking mainly about DNA but not exclusively its a big target and we are looking at test that we do are looking at the changes to DNA and as we say it is a big target and you've got something like three billion sub units of nucleotides in each cell. So there are plenty of things that can go wrong or that could go wrong. So what is genetic toxicology? Well back in the good old days when it first started it was talked about mutagenicity testing but we are not a=only testing for mutations. We are also testing for changes to DNA as well and changes to the genetic apparatus and not all of those will lead to inherited mutations. So we now tend to use this broader term genetic toxicology or genotoxicity testing which not only include mutagenicity testing but it also includes chromosome abnormalities that could be structural or numerical changes. DNA damage affects on DNA repair and damage to the cell division apparatus that can lead to chromosome loss or gain or aueploidy. So why do we do it, well if we look at the pharmaceuticals for example then most pharmaceuticals will be tested at some point in cancer studies or rodent, raddit and mouse cancer studies. Only drugs that ar used for diganostic porcesses or very short

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term use will not need carcinogenicity testing because those tests are time consuming and expensive. Pharmaceutical manufactures want to get some ideas of whether the drug is going to be effective and they want an idea in clinical trials firstly in healthy volunteers and then in patients to see if the drug works. So we need some safety data before we go into human trials or human patients, it would make the drug development process ridiculously long if we had to wait for carcinogenicity data before we could do those clinical trials. So we used genotoxicity data as a surrogate and it is an alternative and gives us a good indication of cancer potential before going into these types of clinical trials. For industrial chemicals like household products, often they are made in such small quantities that they don't require carcinogenicity testing so we don't use genotoxicity data as an indicator of cancer risk without having to do the cancer studies. In cosmetics we cannot do any in vivo testing so we can't do cancer studies, so we have to rely on cancer genotoxicity data to tell us about the safety of cosmetic readings. Agrochemicals in a way similar to pharmaceuticals, most of them will eventually be tested in cancer studies but again the companies developing these want to find something out about how effective they are. They want to be able to do field trials and we use genotoxicity data as a predictor of safety in terms of cancer risk in order to get those trials underway. So the genotoxicity testing is not only about predicting cancer. The examples just mentioned are mainly to produce cancer potential but DNA damage and mutation have other health effects. So although mutation is most often involved in the initiation of tumours. It is also involved in tumor progression so there is a loss or gain of tumor material. There are other conditions such as arteriosclerosis and inborn errors of metabolism that also originate from mutations. In terms of chromosome damage a lot of spontaneous portions and birth defects are associated with chromosomal



damage. So if we look at this below it is a cartoon of colon damage or colon carcinomas. Some cartoons have 5,6,7 steps but this is a 6 step process. Basically what we want to focus on here is there are mutation, DNA modifications this is now in the field of epigenetic. There are alterations of methylation patterns. Activation of oncogenes and then loss or gain of mutation material that might be through

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chromosome deletions or rearrangements or even chromosome loss. So there are mutations involved not just in cancer but in a whole host of other health defects and again as well. In terms of cancer causing genes there are four main types of genes that are involved somewhere in cell division and most tumours have a faulty copy of one or more of these. So oncogenes, tumor suppressor genes, suicide genes and DNA repair genes, the fact that we find mutations in these genes in cancers and tumours is reinforcing the idea that mutations are involved in cancer. So because there are several different types of even involved in cancer and in other genetically derived health problems we need to be able to detect a number of different health changes in any gene toxicity test that we do. We need to be able to detect the mutations structure of the chromosome damage and numerical chromosome damage. But there is no simple test that can detect all of those. So we have to use a battery of tests that complement each other. This means using either two or three in vitro tests. This is usually the first step and the comet assay will be the only step for hazard identification. For many years we have actually used three different tests the latest recommendations now from the UK and the European Safety Authority of minimum factoring that covers all of these end points is an Ames test and an in vitro micronucleus test. The Ames test picks up mutation and the in vitro micronucleus test picks up both structural and numerical chromosome damage. Now because these are in vitro hazard identification tests we really push the limits on these tests where we test with very high concentrations and some very extreme conditions. We also add a rat liver extract called S9 to mimic the metabolism of mammalian livers. The bacteria and cells that we use in culture do not have much in the way of metabolic capability so we need to simulate what will and can happen in the whole human or the whole animal. So we have this metabolising mixture. All of these features the high concentrations or the extreme conditions and unusual metabolising mixtures can lead to artifacts. We need to do this to make sure we are not missing anything important, we are not failing to predict the hazardous chemical but there is a potential for artifacts when we go into these unusual conditions. In terms of David Kurklands work as a consultant is to try to work out whether the results that are being seen are a true reflection of a hazard or are due to the extreme conditions that they are using. We are creating physiological stress abnormal conditions in that they are not necessarily of any hazard. So teasing out the real negative from the false negative and the true positive from the false positive is still a very frequent even and maybe 1/3rd of all the tests that are done will produce an unexpected or an inconsistent pattern of results that needs to be evaluated in some way to find out whether we really are looking at a hazard or whether we are looking at some sort of smoke screen and we are having to deal with it. Once we get past the in vitro test for everything apart from cosmetic ingredients we usually do some follow up in the in vivo tests. Now these are usually hazard characterisations. The first thing we are asking is what ever we saw in the in vitro test, is it reproduced or does it also occur in the whole animal. So for example if we have a positive result in vitro do we see that positive result also in vivo. If the chemical we tested in vitro was negative then by doing an in vivo test we get the opportunity to see if there are any unique in vivo effects. There might be for example a kidney specific metabolite produced and we haven't used kidney S9 in the in vitro tests. There could be reductive metabolism by the gut bacteria which we have not covered in our in vitro tests. So the in vivo gives us the example to see if there are any unique in vivo tests even if the in vitro tests were negative. We now have quite a wide selection of in vivo tests that we can do. Covering the same sort of end points that we could test in vitro. So we can look for the mutations in target genes usually in transgenic animals and we can look at chromosomal damage and numerical damage by

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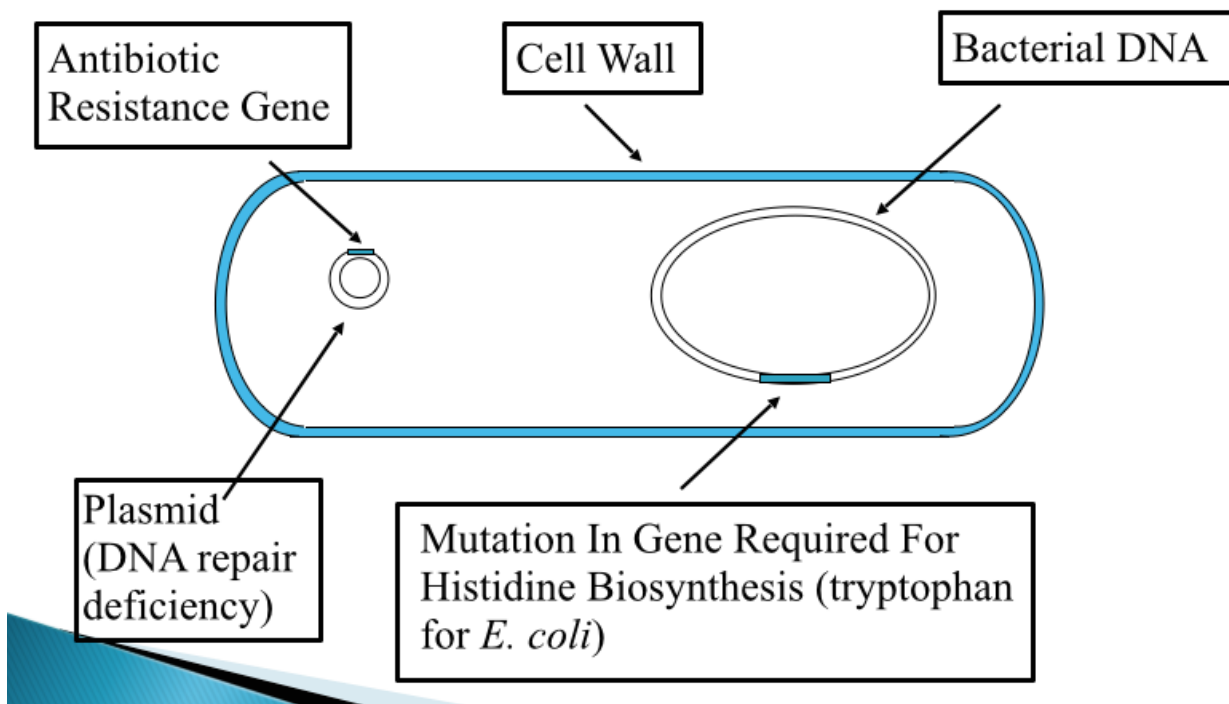
minornuclei and we can do a couple of other things by looking for effects on DNA repair. We can look for effects on DNA strand breakage and we can look for the formation of DNA adducts and we will cover the formation of all of these in these couple of lectures. So in all of these in this first talk we are going to focus on new new patient data and we are going to be talking about three different types of test. Here in the mutation in mammalian cells and mutation in animal models. The in vitro test with the bacterial and the mammalian test are widely used and the bacterial testing in particular. The transgenic mutation test are less widely used because they are very expensive. Transgenic animals and transgenic mice cost something like £200 each per animal so if you are doing the transgenic mutation test you are looking at probably getting on for £100,000 and therefore companies do not undertake those kinds of test unless they have to. So we are going to pause at this point before we describe the test. If heading in to industry then these are the sorts of test that will be carried out and that it is to try and establish whether a compound is safe for human exposure without having any cancer data for example. Then you need to be able to rely on the results. In order to do that we have guidelines which are accepted worldwide to all countries that sign up to OECD (The Organisation for Economic Co-operation and Development (OECD)) There are about 60-70 countries that are part of the OECD and this is to maintain consistency. So that if you test a compound in a lab in China using an OECD guideline then you test that compound in South America then it should give you the same result because you are following a recipe. And trying to follow the recipe for giving those reliable results is what these international guidelines are about and it is what industry has to do in order to produce results that are going to be produced to the regulating authorities. Whether it is the health authorities or the food safety authority, they can rely on it if you follow those protocols. So over the years those protocols have become developed and have in many cases become slightly more intensive and slightly more demanding than they were when they were first introduced back in the 80s. So what we are describing is what we now believe are the best ways to do those tests in order to produce reliable results. The next one is industry contract labs and so on and what they are doing day in and day out in order to report the facilities as being accepted as reliable. Now having said reliable we are pushing the boundaries on these tests and there is the possibility for artefacts.

Ames tests

so this is commonly referred to as a reverse mutation test. So what we are doing is by starting with bacteria that already have a mutation. It is down in one of the pathways associated with amino acid synthesis and in the case of the salmonella bacteria it is used with histamine biosynthesis and in the case of E. coli it is tryptophan what we are looking for is whether by exposure to our test chemical whether we can revert these bacteria from auxotrophic to prototrophic so that they can fully synthesise the amino acid and they don't need the supplement. So we start off having to grow the bacteria in histamine or tryptophan containing medium and then we look for the phenotypic change to the ability to grow in the absence of the additional amino acids. There are several different strains of bacteria that we use. Each of them we use that has a very similar and small genetic target and because the targets are similar in those strains we need to use several bacterial strains in order to cover the different specificities in order to cover the mutagens that we are looking to detect. In addition to this small specific selective target the bacteria

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have been engineered to be more sensitive than a normal bacterium would be by the addition of several other traits. So they can contain DNA repair deficiencies so you can cause damage to DNA and its not repaired its not recognised therefore its more likely to end up as a mutation. The cell wall of the bacteria has been modified increase its permiability to large in partucular chemicals. So they can actually gt into the bacterial cell and actually interact with the DNA. A number of the strains that we use also have plasmids that confer and increased susptability of mutation with out a concomitant sensitivity to toxicity. So this helps us separate the toxic form mutagenic effects in terms of dose range or concentration range of the chemical thats being tested. So just a cartoon of the bacterium seen below.



Theres a mutation somewhere in the histamine or tryptophan biosynthesis pathway. The cel wall has been modified to allow large molecules to enter, some of thes bacteria have this plasmid with the DNA repair mechanism deficeincy incorporated on to it. This biosynthesis gene is simply a marker that allows us to check that the bacteria still has the plasmid if they are resistant to the antibiotic then we know the bacteria has the plasmid. Another aspect of these bacterial tests is that the site at which the mutation has been implemented which in the case of salmonelle is in the HisG HisC or HisD genes. These are hotspot for particular classes of mutagens, in the case of these three markers for GC base base pair or mutations or alterations of the GC base pair. We are able to check what has happened to the DNA sequence around the original mutation by sequencing the DNA in that region and these bottom four shown in the table below here (the HisG46, the HisD6610, the HisD3052 and the HisC3076) are all strains with GC target mutation so these strains will pick up things like base pair substitutions so you have the TC, frameshift mutations with the addition of a couple of cytosines here. Frameshift here with the deletion of a cytosine. TA1537 also picks up a formativet but its more but he way of intercalation. The G428 strain has an AT target so your looking at open mutation and theres quite a wide target and range of changes

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Mutation	Strain	Nature of mutation	Reversion events
hisG428	TA102 TA104	CAA TAA (ochre) (<i>E. coli</i> WP2 also detects ochre mutations)	Transitions, transversions Extragenic suppressors Small deletions
hisG46	TA100 TA1535	CTC CCC	Base-pair substitutions Extragenic suppressors
hisD6610	TA97	ACC-CCT (opal) ACA-CCC-CCC-TGA	Frameshifts
hisD3052	TA98 TA1538	GCC-CGG GCC-GGC (opal)	Frameshifts
hisC3076	TA1537	Not known – presumed +1 near CCC	Frameshifts

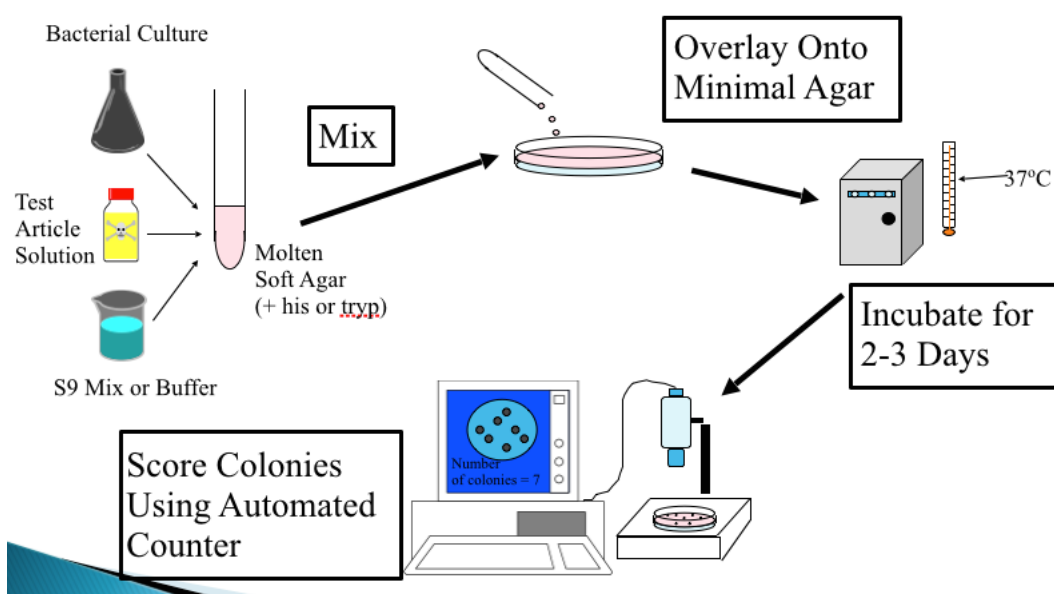
that occur in these. The TA102 or TA104 with transitions or transvertison ETC. So although these targets are quite specific and we know what we expect to be going on when we actually look at the range of changes that occurs in human bacteria. So for example if we take TA102 the HisG428 then we actually find that tater ar multiple changes that have occurred in order to achieve this reverse mutation. so we see a true reversion which is a simple base change from aT to GC. But we also see the inorganic suppressors have been mutated or there have been large deletions with 3-6 base pari deletions and we can also get mutations in extragenic supressors. So although these are supposed to be quite engineered to be very specific and very selective mutation that would probably only be reverted by. One single event we actually see multiple events particularly int eh HisG428 strains with these ochree mutation. We mentioned metabolic invasion and of course bacteria don't have livers. In humans your exposed to a strange chemical and the liver sees it as not being particularly useful and tries to preparee it so that it can be excreated and that means that in the liver these compounds are metabolised and particularly in the liver it tries to make them water soluable and if you have got an organic chemical and you have ingested an organic chemical, your liver want to make it water soluble so that it can be excreted in the urine to get rid off it. Now that metabolism is generally oxidation metabolism. sometimes it makes a mistake so if you breath in bozo-a-pyrene or if you swallow bozo-a-pyrene for example your having a barbecue, having a bonfire, eating the same time. You take in some bozo-a-pyrene and your body docent like it and tries to get rid of it so it metabolises it and wants to make it water solublee by adding hydroxy groups. In the process it makes an epoxide and that epoxyde binds to DNA and that is what is mutagenic and carcinogenic from bozo-a-pyrene. So in the process of our own livers trying to detectively our bodies form that chemical it actually makes an intermediate that binds to DNA and causes mutations. The body is only trying to do the right thing by removing the chemical but in the process of doing s there is a DNA reactive intermediate or a idol-epoxide which binds to DNA and causes the mutations. So it is important that we include some aspect of metabolism in to the testing. So we take some rats and pretreat them

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with this either the polychlorinated biphenyl or a mixture of phenobarbital. These substances stimulate the liver to increase the enzyme level some of the p450s and in particular p450s involved with oxidative transformation. So the S9 is a 9000 G supernatant from the livers of the rats that have been pre-treated with this compound. This supernate contains mainly high levels of mono-oxygenases, oxidases and aminidases etc. Some of these p450s are amplified 40 times the level that you would find in a normal liver. so they really are hyped up to try to push any metabolism in identifying the hazard. Now the S9 wont do the work on its own, you need an energy source and you need co-factors so we have to add those into he mix as well. So when we are going an experiment we are growing batchria overnight where they are unusually diluted so there inactive growth because our test chemical. We have our S9 and co-factors or we also do the treatment in the absence of the metabolism if there are any direct effects. So for example an alkylating agent like methylmethol sulphanate docent need a metabolism. It will cause mutation without metabolism so we will pick that up better if we pick it up in the absence the S9 mix. So we do it with an without the S9 mix and there are two ways to do the test, we can mix all of these together in a coupe of MLs of our soft agar. Mix it all together and pour it on a plate and apply the colonies. Or we can mix these together without agar for about a hour half an hour maybe and then add the agar and do the plating. So without the agar its called pre incubation and of course what this dose is it you can have much high concentrations of both test chemical and the liquidd S9 and the bacteria. You also have a completely liquid matrix, so you increase the chances that the compound can get into the bacteria or that a metabolite can ge tinto the bacteria but you also increase the chances of toxic effects. So there are some advantages to doing pre incubation but there are also some downsides to doing pre incubation where it can be very toxic. Now you will see this little bit here in the slide below that in this cocktail we have a small amount of histadine or tryptophan. Can anydy tell me why they think we need to have that? We are starting with bacteria that are deficient in the ability to synthesis histamine or tryptuophan so we need to make sure that those bacteriaia go through a few cell divisions cause any damage to the DNA that

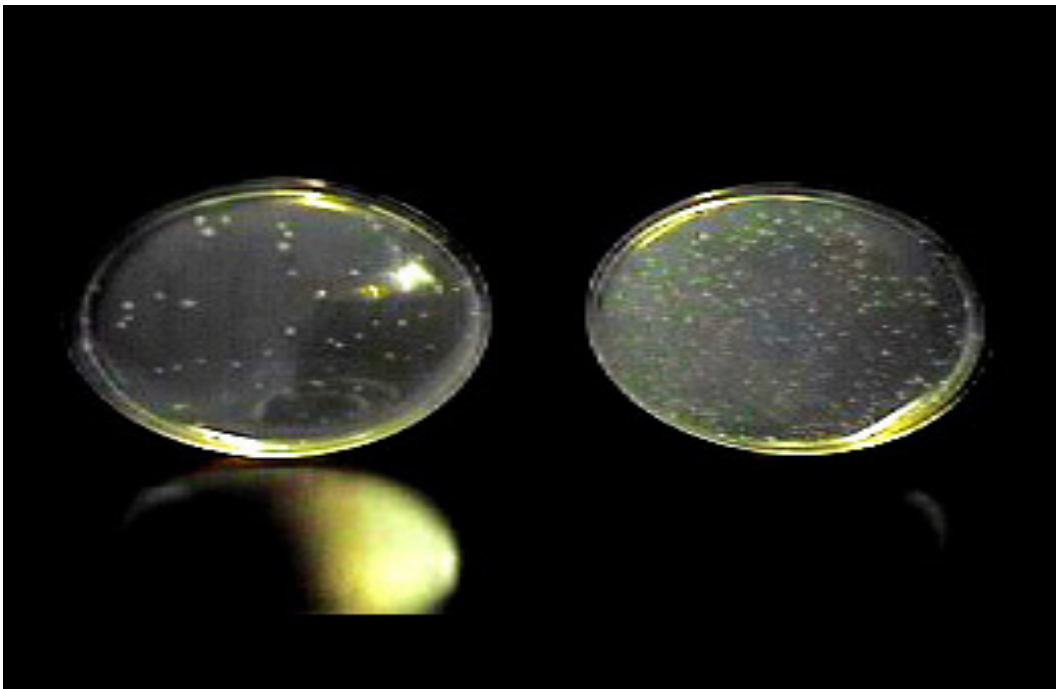
our test chemical has caused needs to be fixed. So it needs to be in a permanent state or a reliable state where the changes tot eh DNA need to be consistence and persistent and that change then needs to be decoded into the various enzymes that are needed for the histamine biosynthesis. So the damage to the DNA needs to be consistent and the RNAs

The Ames Test: Basic Assay Design



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and proteins that are down stream from that DNA need to have had time to change. So you need to give the bacteria time to get from a mutation that means it can't produce histamine or tryptophan to a native DNA that can produce histidine and that means you need the bacteria have to go through a few divisions. So in order for them to make that happen we need to include a little bit of histidine or tryptophan in this mixture. Now it is only a little bit and when the bacteria have used it all up which they will quite quickly then only the mutant bacteria will grow. The starter bacteria that have not mutated can't grow because they have run out of the little bit of histidine or tryptophan that we put into the cocktail. That's important as we will see in a couple of minutes. So these bacteria, we normally put about 10^{12} bacteria on a plate and because they are engineered to be so sensitive then even on a control plate we get a number of spontaneous mutations. So this jazy are in the background are the starter bacteria that can't grow in the presence of histidine, they have gone through a few divisions in the presence of histidine



that we have include and then that has caused them to stop growing. So this is a lawn of micro colonies. So they are starter bacteria they have not mutated they have gone through a few division and then they have stopped dividing. Only those that have mutated and are now independent of histidine that can synthesis their own histidine to grow on to form discrete colonies that we can count. so this is a part treated with a positive control chemical and as you can see there are a lot more colonies and a lot more mutants. So it just becomes then a numbers game. We need to treat enough cultures we need to treat enough concentrations and we need to treat a number of replicates. We won't dwell on it, this is just a summary for easy reference if you wanted to look at it. Because the different strains have different types of mutation we are going to cover every possible type of chemical damage to DNA. Then we need to use a number of different strains and usually there are four strains of salmonella with a GC target so these strains are

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- TA1535
- TA1537
- TA98
- TA100

one or more strains that can detect mutations acting at AT sites so this would either be

- TA102

or one of the E.coli strains like

- WP2 *uvrA*
 - WP2 *uvrA* pKM101
-
- ▶ Need basic 4 strains of *Salmonella typhimurium* (G-C sites)
 - TA1535, TA1537 (or 97 or 97a), TA98, TA100
 - ▶ Plus 1 or more strains to detect mutagens acting at A-T rich sites
 - Either TA102 or *E. coli* WP2 *uvrA* or *E. coli* WP2 *uvrA* pKM101
 - ▶ Each strain detects different effect
 - +ve in only 1 strain indicates hazard

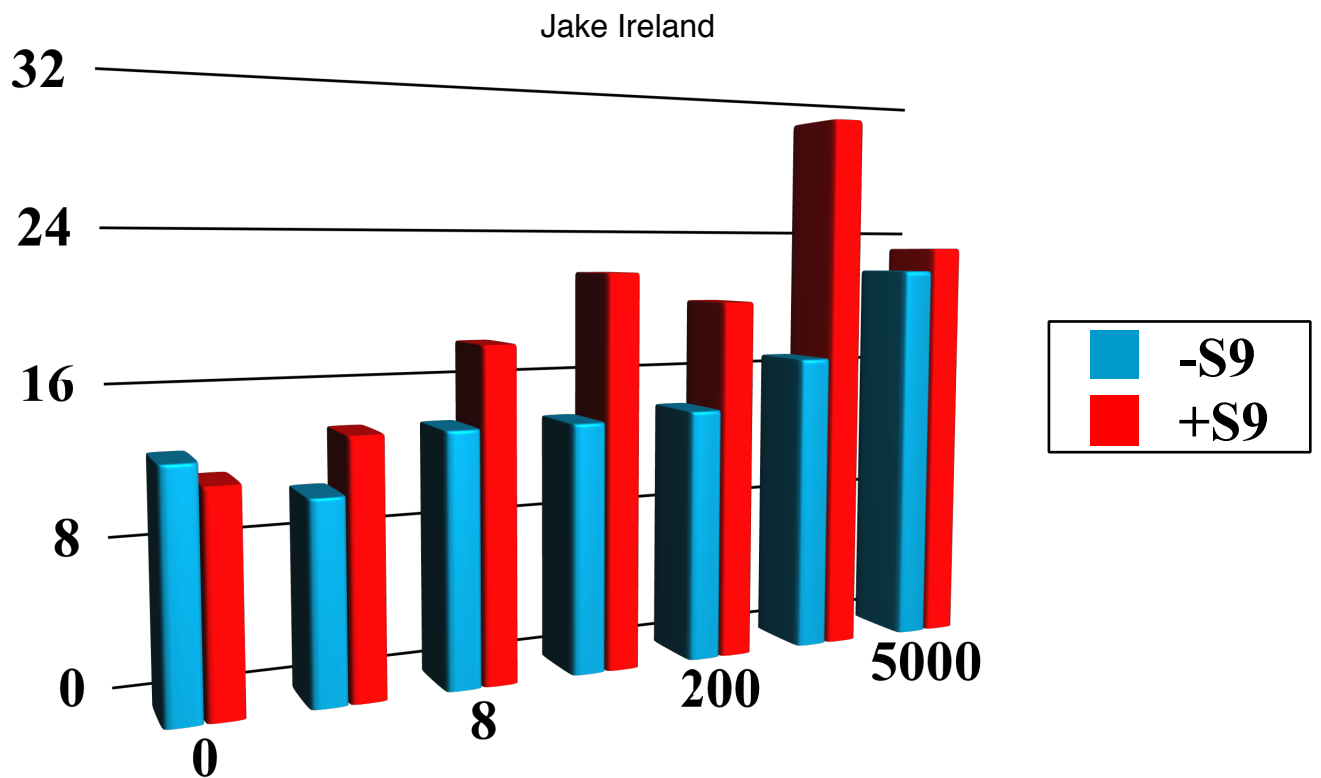
And because each strain detects a different effect we only need a positive result in one strain to indicate a hazard. We often getting nice clients saying well we tested it in 5 strains and four of them were negative so that outweighs the positive well no this is not scored one strain that's positive is telling you something important and we are not looking at dull efforts. So its important that plate incorporation and pre-incubation are both available. We mentioned there are pros and cons for each but from an ideal scientific point of view you would actually do a test using both methods but now many of the guiltiness tell us that we only need to choose one of these and that will be ok.

As we mentioned earlier we do go to extreme conditions with testing in 5mg of platelet which is quite a lot of chemical. We can test insoluble concentration, we can test some compounds that precipitate as long as it doesn't interfere with the story and this is quite useful because it can allow us to detect impurities. It is very difficult to synthesis a product and make it 100% pure technical materials are quite common levels of impurities are quite common and they may be mutagenic. It is helpful from a health and safety point of view to know that that's the case. Generally in the ames test we do three replicates per concentration and five concentrations and we do the test with a n without these metabolising agents. Now as we have said if it was an ideal world then we would suggest that if you do an experiment then you should repeat it because scientific rigour suggests that results need to be confirmed and this would be needed to change the conditions to do both plate incorporation and pre incubation but for human pharmaceuticals this is now not needed a single robust experiment that is clearly positive or clearly

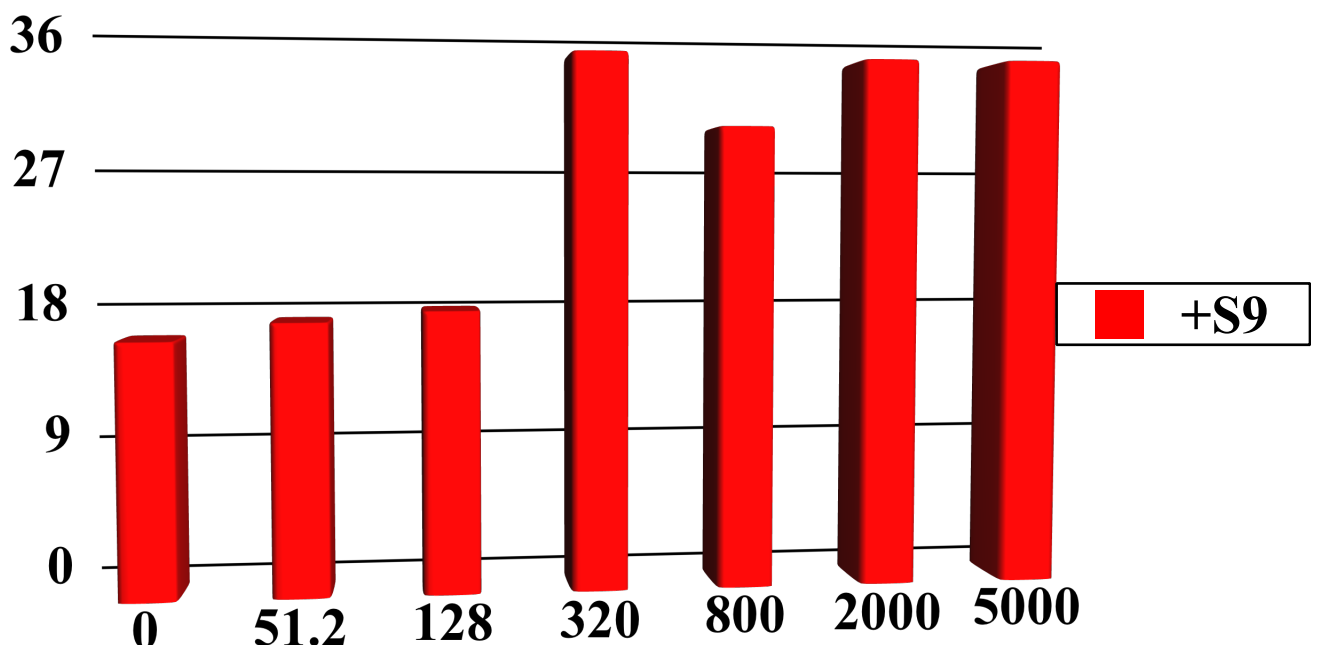
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negative dose not need to be confirmed. If you do do a second experiment the best thing to do is not an identical repeat but to change the conditions like the pre incubation to plate incorporation or the concentrations of the test chemical or to change the concentration of the S9. As long as you have fourfold all of the parameters that are in the guideline with enough replicates and enough dosing then a negative control and positive control and full accounts of the colonies then should be an acceptable complete experiment. Ames test have been around since 1973-4 and is still at heart of all regulatory process. People say bacteria don't get cancer but these are very good indicators of cancer potential.

We said that we need to add a bit of an amino acid to get the bacteria through a few divisions with the mutation. Fix it in the DNA and allow then for the downstream RNA and protein changes to occur. So basically just allow the bacteria to synthesis histidine or tryptophan for themselves. What do you think happen if the substance that we are testing contain one of those amino acids? What if we are testing a plant extract that contains amino acids or the pharmaceutical industry is synthesising a peptide to treat maybe some kind of genetic disorder and that peptide contains some histidine or tryptophan, what's going to happen? It will basically grow on the treated plates where we have got more histidine or tryptophan the bacteria will go through more divisions. What's that going to lead to? It can then revert back to its original mutant. First of all if you get a lot of growth of bacteria then that background lawn of growth is going to be vast and you won't be able to see the mutant colonies. Before you get to that point, the bacteria on the treated plates so with the amino acids will go through more divisions before that's used up. For each time a bacterium divides, there is a specific chance that it will mutate so the more divisions it goes through the more spontaneous mutations it will occur. So you end up with what looks like a dose response where you end up with increasing numbers of revertant colonies on the plate that is just due to feeding. So the histidine is allowing the bacteria on each of those treated plates to go through more division before it's used up and with each division there's a chance of a spontaneous mutation so it's not getting mutation due to interactions of the test chemical with the DNA it is simply feeding the bacterium. which is increasing the frequency of spontaneous mutations and you can fiddle and change the way you do the experiment. you can do a treatment placement experiment. Whereby after treatment it would be the preparation that might contain the amino acid. You can spin and wash and take the test chemical out of the test chemical before you put it on the plate and then it can't feed the bacteria. So you can reduce the risk of basically getting a false positive because it's not a mutagen it's a food source. People particularly make products from plant extracts and of course in the chemical industry it makes oils and fats and proteins from plants that are supposed to be good for your skin and your hair and that sort of thing. They can cause problems in this kind of test. So some audience participation. This is a test that was done a few years ago with strain TA1537 that just using plate incorporation methodologies the red bars is with S9 and the blue bars without S9 where we got up to the extreme of 5000ug. seen below we at that time used to do statistics quite regularly and this distinction in the Ames test can pick up increases of less than two fold that has been statistically significant so this was the first experiment we did and if you were doing this in the lab and you had to go tell your boss down the corridor that the Ames test got this result. What would you be telling him? Is it positive or negative not sure? It looks positive because you certainly wouldn't be telling him it looks compliantly negative so we did the right thing by doing the experiment



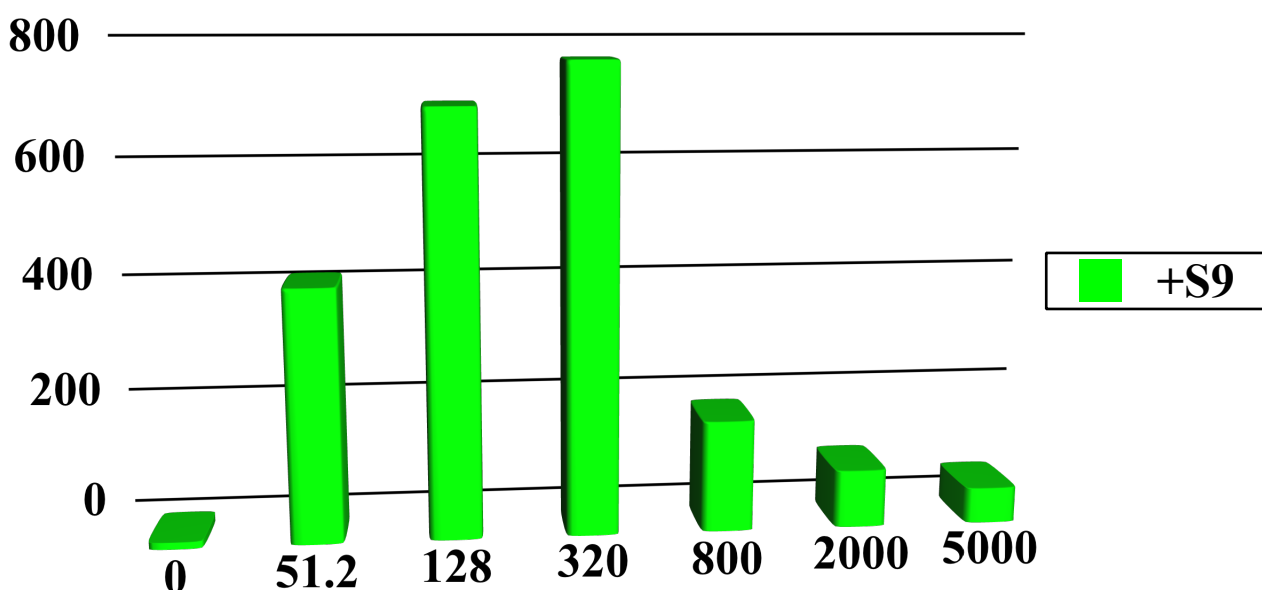
again and we changed the dose range. So we closed in on the area that the dose range that we saw something and it was reproducible and now instead of doing something at one dose point we have closed up the dose range this is the diagram seen below. These are all statistically significant with all about twice the background. So negative positive? it is reproduced so it looks positive but what we have to think



about is the general rule of thumb for this strain is that you only accept a threefold increase as being biologically significant. Now that is because the spontaneous counts with this strain can be anywhere between five and fifteen or five and twenty. So you can get a three fold differences just in your control

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plants and therefore tend not to accept that a response in that strain becomes biologically significant unless it's gone up more than three fold. So this result would not be positive even though it's reproducible and even though it's not a clear negative. What would you do with something like this? We have done plate incorporation. This should be made clearer that this strain is specific to the effects of this compound where all other strains were negative and this one was the only one that saw an increase and it was only present in the S9 positive tests. So we could play with the S9 concentrations or we could try a different S9 product. We could do pre incubation now then. so these results are what are called weak positives and it is less than threefold, it's not however clearly negative but we wouldn't however get overly excited about it. Now we did a pre incubation experiment and we got this graph below.



So we are at hundreds and hundreds of mutations and fold increase in the mutations just by doing pre incubation and it's because of results like this that we should worry when the regulators say well we can choose pre incubation or plate incorporation it does matter pick one and stick with it. This is an example of a stark difference when the difference between the mutagenic activity between the mutagenic activity between - and + S9. This then was explainable where this compound was an anthraquinone. Now anthraquinone are flat molecules or planar molecules and they can easily slide between the base pairs in the DNA and that's what TA1537 detects it detects a frameshift mutation by a chemical slotting in between the base pairs and distorting the helix and that's what this was doing after metabolism. It needed metabolism, it probably needed one or two of the side groups to be knocked off so that it could be slotted into the base pairs. Now unfortunately we are in the contract industry and we often don't get to follow up on these compounds so we don't think this compound was ever progressed and we don't think this compound was ever tested for carcinogenicity. We don't know whether that strong positive Ames test was an indicator of cancer potential because it never got that far and that's one of the shames because sometimes it gets some really interesting results and you don't get to follow them through so you don't get any significance of it. But for me that was a stark example of how if we had only done the plate

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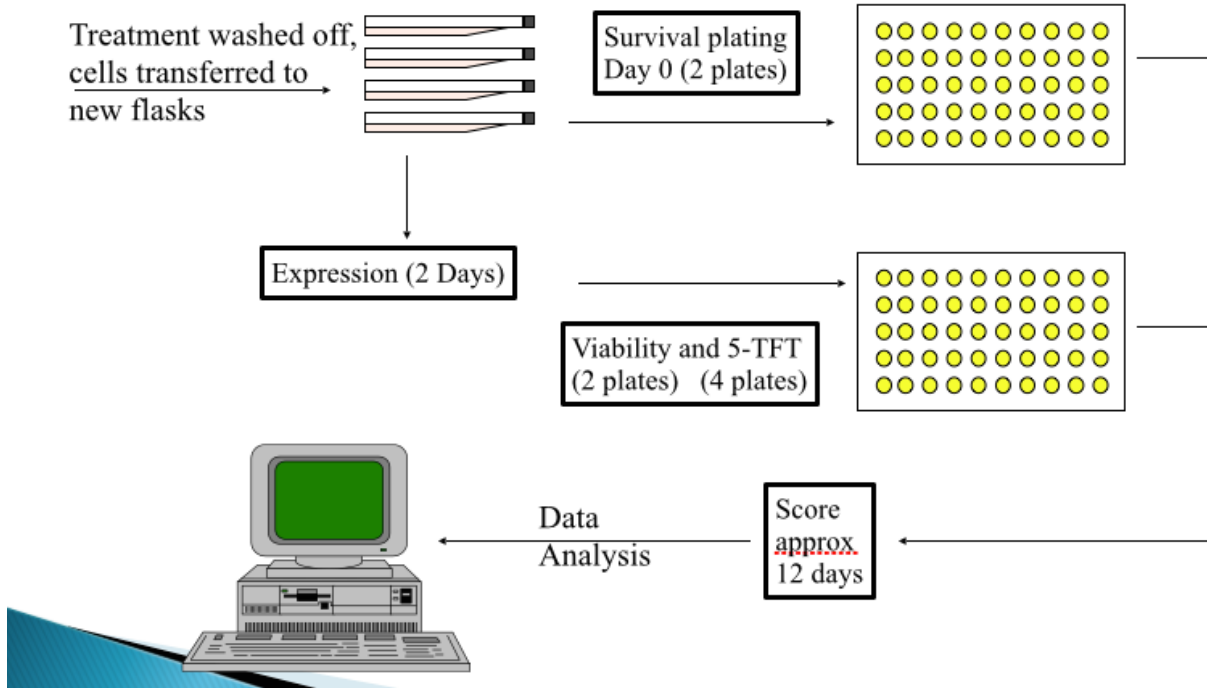
incorporation test then we would probably have said this is pretty weak, that might be due to an impurity. Lets synthesis another batch and do our in vivo test and mammalian cell and it will probably be negative and it would have got swept under the carpet. And we never knew whether that really strong pre incubation response was a true indicator of a real problem.

So thats all on the ames test and now we are going to do mammalian cell mutation and transgenic mutation. Now we can see things happening in bacteria that may not happen in mammalian cells the organisation of the DNA for example is completely differne . The bacteria don't have sets of chromosome that divide in the same way they do in eukaryotic cells so you have got a spinal apparatus and poles controls that occurring mammalian cels that don't occur in bacteria so we couldn't rely whole on a testing bacterium so we need to have a look in amamlian cells and one of the most common mutation tests is in the mouse lymphoma tests were it is looking at the thymidine kinase. Basically we start with a heterozygote plus minus thymidine kinase and what we are looking for is whether treatment with a chemical can treat that to TK minus. So the starting bacteria can synthesis TK the mutant bacteria cannot synthesis TK so if we put in a elective agent like triflurothymidine which thymine kinase enzyme will convert to a lethal form then we a have a simple genetic system in the presence of TFT the starter cells die and the mutant cells service so it a simple selective system. Now this si different from the ames test where thames test was a conversant mutation and as we described in one of the earlier slides you can see some very specific mainly point mutations and occasionally simple frameshift or a single base frameshift or deletion. Because the mouse lymphoma assay is a forward mutation system it is not a reverse mutation system. It can detect a much wider range of genetic changes. So not just point mutation but intragenic deletions and allelic deletions, deletes associated with non-disjunction and recombination mutations and this shopping list is the range of genetic alterations that we find in tutors cells. So other people that say well id actually do this assay is much more relavent at predicting the changes that occur through forming a cancer cell because of the range of different genetic changes that it can pick up. To compare with the ames test be course this maximum somethignsystem can detect such a wide range of genetic events it has a higher spontaneouse mutation frequency. Theres more opportunities for spontaneous mutations to occur. So we don't need to treat as many cells, where as we normally treat about 100 million or 10^8 bacteria on an ames test plate. We only need to treat a few million mouse lymphoma cells at this concentration because of the spontaneous concentration inside. but one other important thing is is that these mouse lymphoma cells just like the ames test you have to allow the bacteria to grow long enough for the reverse mutation to allow the cells to synthesis histamine in the case of mouse lymphoma cells we have to allow the cells to grow long enough to get rid of the thymidine kinase. So those TK + - heterzygotes they start with some TK in the cell or within the cytoplasm. We hit them with a chemical and mutate them to TK - - homozygotes but that TK is still there. There not synthesising any new TK but they have go get rid of what is already there otherwise those cells will die in the presence of the selective agents. So we have to go through whats called an expression period when the TK - - genotype becomes fully expressed and there is no TK within the cells. Now that expression period requires several divisions a bit like the bacteria going through several division in the presence of histidine. That growth period between the initial DNA adamage which is the genotypic change and when we select for the phenotypic stage thats called the expression period. Now in bacteria thats very short because bacteria divide every

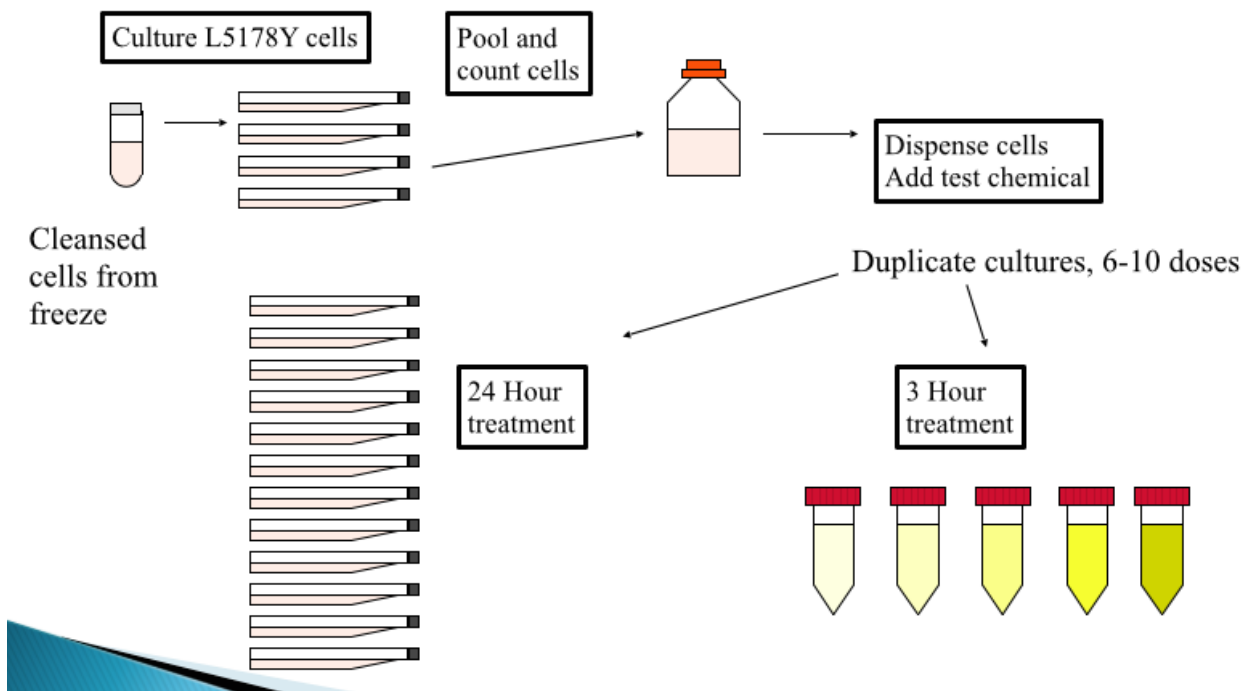
Jake Ireland

20 minutes. So that expression period is very short. What's the expression time for a mammalian cell like this then? There's about 15 hrs. So we have to go through a couple of days for the genotypic change to be expressed as a phenotypic change. Now because we have to subculture the cells during that period we have to check that they don't get too dense and that the cultures don't stagnate and then we have to get into the selective medium and they have to grow in that. So the cell numbers are changing during this period. So we don't need to correct or calculate the immune frequency in the Ames test we just count colonies on a plate. But here we are going to have to relate the mutant frequency or calculate the mutant frequency based on the viability. Because the cell numbers are changing and the whole growth period is much longer than it is in an Ames test. So that's another difference. So again we test to high concentrations to 10 milli molar or 2mg per ml. We can't really handle ppt in these of cultures and it really does get in the way because you are subculturing and these cells grow in suspension, they don't grow as a monolayer on the plate. They have to grow in suspension. So washing out a ppt is really difficult. So we have to avoid ppt conditions but we need to know if we have gone high enough so normally we will include one insoluble concentration. For human pharmaceuticals they actually lower the top concentration so it's different from testing all other chemicals. We usually test four concentrations. We prefer duplicate treatments but there are labs that do single treatments and maybe they'll test more concentrations but again these cells don't have much in the ways of metabolic capabilities so we need to test with and without rat liver S9. But in mammalian cells you can't treat for long periods with S9, this is because those oxygenases or the oxidative enzyme capacity is a problem. If you leave that in the presence of the cells for too long then you get break down of the lipid membranes and that produces reactive oxygen species that is toxic and causes DNA damage. So we can't treat for any longer than about 6 hrs in the presence of S9. It just becomes too toxic. So basically the protocol is to test for 6 hrs with and without S9. We used to also include an additional longer 24hrs treatment in the absence of S9 but the latest recommendations are going to go before the OECD coordinators in about three weeks time and they are suggesting that that is no longer needed and that that 24hrs treatment is no longer needed. There are some people that are worried about that and think that it should be included as it makes for a more supportive test. So we are going to have to see if that's going to be approved or not. Because you're doing subcultures in the cells because the cells are toxic so you'll reduce the number of cells you have to engineer the test to make sure that at every step and at treatment through expression you are retaining enough cells to give at least 10 and preferably 100 mutants. That's to make sure you avoid zeros, we do not like zeros in these experiments. So if you had zero mutants on your control plates or something like that it really doesn't give you any indication of the sensitivity of the test or its ability to detect a mutation. So by making sure that we subculture and carry through enough cells to always contain the defined number of mutants then we make sure we avoid the zeros. We particularly want to avoid zeros on the control plates and if we can we want to avoid zeros on the treated plates because that might mean that there are so few surviving cells that we are missing or failing to detect the compound's effect.

MLA: Summary of Practical Procedure

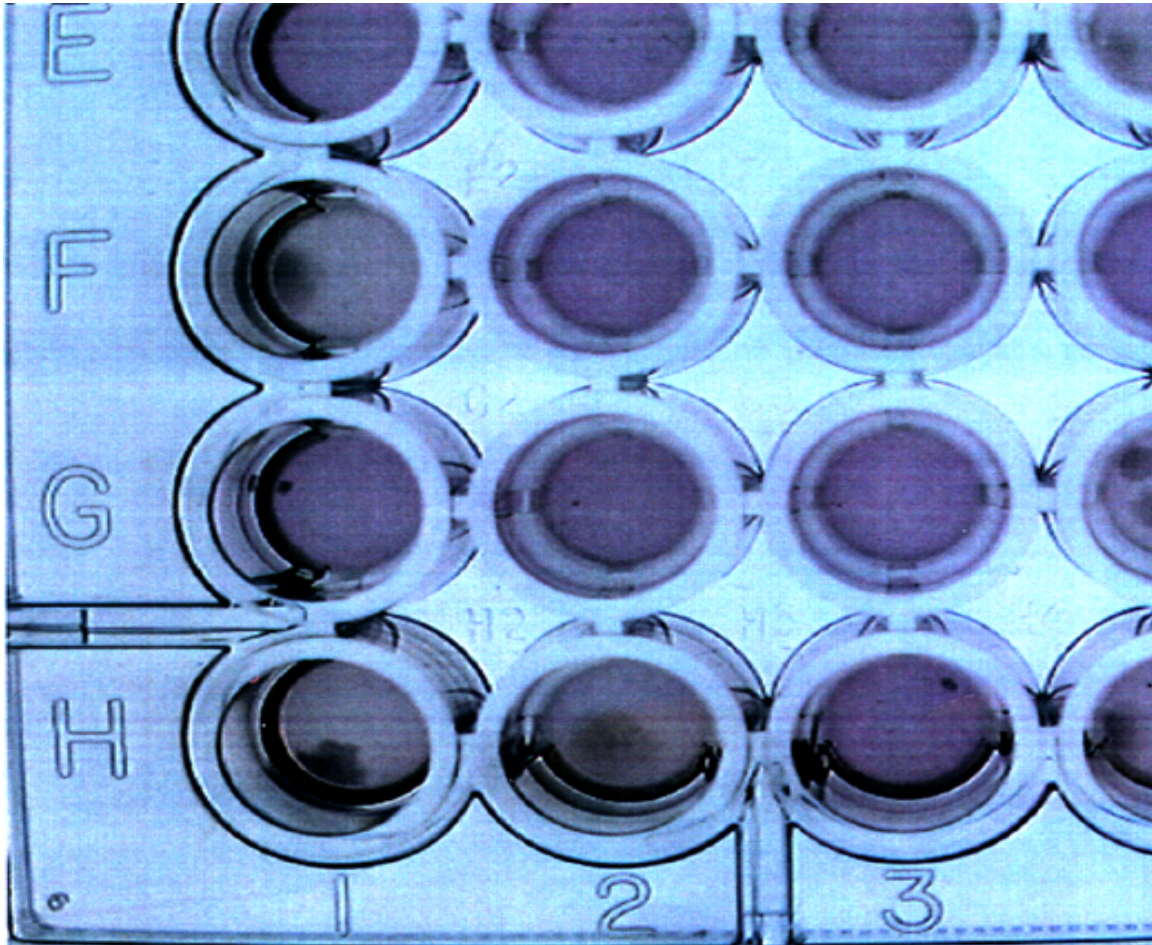


MLA: Summary of Practical Procedure



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So just a couple of cartoons to for who this is conducted. we need to rapidly grow cells and they need to be cleansed pre existing mutants before we grow the cells. We grow them and count them and in short treatments we put them in tubes as because its only going to be for three hours if we are doing the mon treatments we put them in flasks and then after the treatment and wash off we express the two days then we count. We then plate for survival and we place like two cells per well and we also plate for in selection medium mutation plates where we will be plating them out 10^5 cells per well and these are then incubated for a bout fie days to allow the colonies to grow. Now we can do this, we used to always to this in the 96 well plates. But you can do this in petri dishes or in agar and in both cases you get, two different



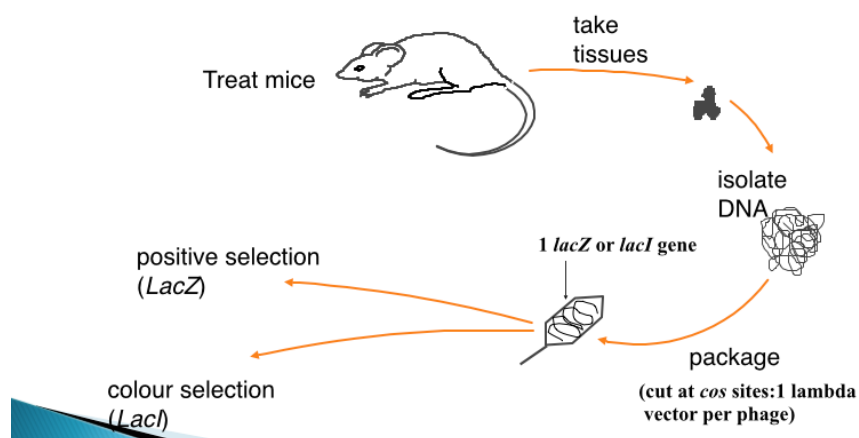
types of mutants. You get some wells will contain a colony that is quite pale and maybe occupies a third to a half of the well after 12 days incubation and then in other wells you get small compact colonies. Now these have been analysed genticly and basically these larger colonies have quite small genetic changes or point mutation gene mutations or very small deletions. Techie are able to grow at a normal rate and normally dividing cells, whereas these cells that form the small colonies are firstly much slower growing which is why you have a small colony and they tend to contain much larger genetic damage so larger genetic region or rearrangements. So the types of colonies that we get can tell us something about whether not we are looking at a point mutation or a chromosomal change or chromosomal damage and that can be quite useful. So thats the mouse lymphoma assay, which is quite different from the ames test but ti dose give you different information.

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So to finish of this first talk we are going to look at transgenic animals, now these have been around for a while but we have only had an OECD guideline on this test for about three or four years. It took while to get enough data together and as we mentioned earlier these animals are very expensive so you don't tend to get a lot of people publishing data on them. The two most commonly used transgenic animals are both have a target transgene associated with beta galacisade activity. The the case of LacZ that is the beta galactosidase gene that in the case Lac I this is the promotor for the beta galactosidase gene and these transgress are located on a lambda bacteriophage vector which has been incorporated into he mice and there are multiple copies, in the case of LacZ ts 40 copies. concatenated head to tail in one location. So there are multiple copies of the transgene and again you have quite a large target. One interesting thin is that the transgene is neutral, so if the transgnee is mutated by treating these mice with a chemical it disant confer any advantage or disadvantage on the animal and that means there is no selection pressure. There is not selected for or against and its quite neutral and that is quite an advantage because it means you are looking at real absolute effects when looking at mutations in that transgene. So what we do is we treat the mice and we will go through this protocol in a minute. We extract DNA now the lambda bacteriophage vector has enough DNA there that when you take the DNA out of the mice you can add to it whats called a packaging mixture and this is a mixture of enzymes and portions that allows the lambda bacteriophage to reconstruct it self. So it basically takes all the bits. We add the enzymes and proteins and things like that and the DNA does the rest. So we put this stuff together and we get reliable lambda bacteriophage. Now those lambda bacteriophage will contain the transgene. Some of which will be normal others of which will be mutated. We then test those bacteriophage on a sensitive strain of baitera. Its usually E. coli C and the bacteriophage of course kills the bacteria and forms plaques or clear areas on the plate. Depending on wether that transgene is normal i.e. expresses beta galactosidase or is mutant i.e. dosnet express beta galactosiadase then we can see the mutants form the non mutants by a colour reaction in the agar plates (blue white screening) So here is a cartoon of the transgene of a lambda transgene or the lacZ transgnee on the lambda bacteriophage vector. At the end of each insert are what are called Cos sites. These are important because they need to be intact when we extract the DNA and add it to the

packaging mix, if those Cos sites have been destroyed then the lambda bacteriophage cannot constitutete or cannot reform. So if we are treating the animals with a test chemical that causes large deletions and it deletes through those cos sites then we may =will have caused a mutation but we can t detect it because we cant get viable lambda bacteriophage out. So this test system is not very sensitive to chemicals that cause large deletions. It is very good at picking up chemicals that cause point mutation but chemicals that cause large

MUTATION IN TRANSGENES



Jake Ireland

DNA deletions we might get Cos negative. So just to go through it again, once we have reformed our bacteriophage they will lyse E. coli sensitive bacterial called to form plaques. We grow those on an agar containing a substrate that releases a colour or a blue colour if it is present. So mutant plaques form a blue colour and wild type plaques form a colourless so there easy to count and with the LacZ system its easy to count and use whats called a positive selection system which means that the wild type bacteriophage die before they can form a plaque and therefr we are ony scoring mutant plaques and that makes the whole experiment much more methodical because we need much fewer plates and equipment in the way of incubators and things like that. So its basically an ex vivo assay and this is just to show you that the mouse looks like a normal mouse it docent have two heads of anything. AAlthough the mouse is widley used and widely published systems are MutaMouse and Big Blue there is quite an interesting model that has been developed in japan that combines GPTdelta that is another enzyme based locus with a spinal mutation. This can detect larger deletions as it is not so susceptible to trans negatives and importantly there is also a rat model, so these are mainly a mouse system and of course rats are quite useful because in most industriess you have much more data in rats. So you know about the toxicity the levels of blood and things like that. So getting superting data is somewhat wasier than if you have got a rat model. There eis much less data published in that. Now some chemicals induce mutations quite quickly in these in vivo models and others take quite a while to get built up and because these are neutral mutations they don't disapear. So once you have gotten to a neutral platoit docent matter wether it takes three days of fifteen days. Once you are at a plato it stays there. So we don't have to worry about loosing mutants and therefore we can treat for a longer period so to cover all of the bases we generally treat fro 28 days and then sample about three days later. So faster acting mutants will produce mutations and then they will plato earlier but other compounds that need metabolism will have there effects later. The germ cell mutagens shows there is a suggestion now that we might now need to go even longer. We can detect mutations in early stage germ cels at 28 days but if we want to detect mutations in sperm for example we probably need to go out to 56 days or even longer. So this protocol might change in detecting mutations in germ tissue. so these atre quite expensive expressents, when your looking at £200 per mouse and you have y=to dose for 28 days and you have to take lots of DNA sample and do all of the ex vivo bacteriophage manipulations and so on you see how they can get expensive and time consuming experiments. But they do provide some quite valuable data. You need several dose levels in order to look for a dose response. You need enough animals per set per group in order to look for heterogeneity but the useful hing about these models is you can datect mutations in and tissue. and we mention that now because when we go to the other talk and walkabout one of the other in vivo test that have been widley used is much more limiting for us to be able to look for mutagenic or genotoxic effects in any tissue at site of the contact for example or GI tract or lungs. As well as liver kidney brainn whatver is it really useful. So fairly strait ofrwardfrom the technical point of view and only hard due to money and time, treat the animals or take the tissue. We can put those tissues in the freezer, we can take or we might say we are interested in the liver but we better take a bit of GI tract and a bit of kidney as well just in case we need it later on. We can put those tissues in the freezer. You can store these for up to five years and then still just extract the DNA from them mix with the packaging mix and then do the ex vivo mutation expertments. So isolate the DNA add the packaging mix get that in the bacteriophage and plate on to to E. coli C and use the positive selection system in mute mouse to reduce the number of plates or a

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recover selection system with Lac I. So that's the transgenic mutation models. So yes just because it causes a reaction in a mouse or a rat doesn't mean it will be the same in a human but if you compare rats with mice so let's say you are concerned about the ability of a chemical to cause a cancer in the rat so you did your test in the mouse. The mouse then trying to predict what happens in the rat ok. That is only accurate to 70% and that's between rats and mice. So the difference between rodent and humans is probably going to be even bigger. Now history tells us that there are chemicals that we believe are carcinogenic to humans and there are organisations like IARC (the international agency for Research on Cancer) lists 106 chemicals that are considered to be human carcinogens. They are all rodent carcinogens so the rodent isn't missing anything that we are aware of that would be a human hazard but it has been probably been giving positive results with a lot of chemicals that are not human carcinogens. Meaning that they are over predicting. The regulators and health authorities would rather have it that way round though. They would rather the animals be more sensitive than the humans because then you not letting anything through the net. It causes problems for industry because it means they could be getting positive results in rodent cancer studies that are not indicative of a human and if they want to progress with that compound then they are going to have to do a lot of work to demonstrate that it's not relative for humans. So for sure there are differences but if you like the differences are on the safe side. There has been a lot of follow up work on Chernobyl and in Japan in Nagasaki and Hiroshima for example. For sure there were increased frequencies of leukaemia and thyroid cancer due to increased radioactive exposure and there were reproductive effects like fertility and so on but we have not heard anybody speak on this in quite a while but the impression that was given is that it is not as dramatic as everybody hears. Humans are pretty good at recognising damage and in particular in the reproductive process. If a foetus is damaged then humans are good at recognising it before that foetus develops. And spontaneous abortion within the first month is normally the result of that and then we are unaware that there was anything abnormal embryo. So yes there are lots of things to consider and humans are pretty resistant to a lot of these environmental exposures. Then having said that we worry about the fact that there seem to be so many more allergies now than 40 years ago. Is that because we have been exposed to so many synthetic chemicals that what we used to be? is that more of an issue in terms of health in terms of cancer or genetic disease or is genetics involved. Is it genetics that leading to the increased number of allergies. Is it up to the pharmaceuticals to continue with the progression of a test even if they get a positive result in one of these tests? usually it's going to depend on three things. 1) do they have a backup compound that is going to be safe? Big pharma will usually be looking at a bunch of chemicals all at once that have subtle differences, so they may have a back up where a small company may not. 2) what are the economics is this drug going to treat a large number of patients or a wide spread population so that the returns are going to worth the additional effort to resolve the extra effort to look for ways around the deskmating results, if your looking at a very small patient population then your not going to sell them any until of the drug then it's probably not going to be economical. 3) what is the medical need and are there any effective treatments already out there, because the regulator may well accept a number of unanswered safety questions if the medical need is dramatic enough. So for example in the last year is if someone had come along with an untested Ebola cure then people would have said well there's a 70% chance these people are going to die we don't care whether it might give them a mutation. If it's going to save there lives now then we will try it. This is the situation that we were in with AIDS drugs a number of years

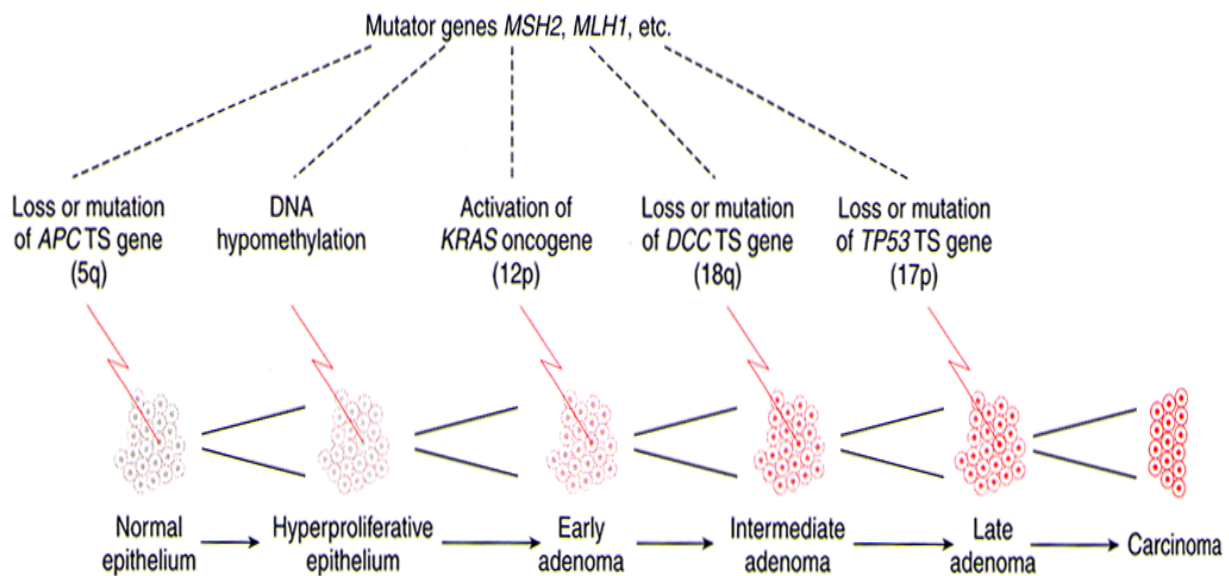
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ago where the regulators they didn't care or not if those treatments caused cancer or not and many of them did because the nucleoside analogs which are antivirals did cause mutations. Now because so many people are surviving aids and the treatment of aid sand there living longer. Now the health authorities are concerned about the areatments for aids and wether or not the drugs that treat aids might subsequently cause mutations. So it wasn't a cause for concern many years ago because it was a considerably un met need. It now is a concern because people ar living the disease longer. So the consideration you have to take into account is where each one needs to be taken case by case and askd these questions of do we have a back up? who do the economics look? and what is the medical need? thats what your going to cause your dissicion on? Is it worth it or isn't it? generally the pahrmaceutical companies will make these dissections themselves because they look much better and they are seen in a much better light if they go to the regaltor authorities with a coherant story. So you don't want to go to the regulators with these results and say we don't know what to do about it, because thats not a strong place to be in and the regulators might tell you that they want you to change the world before they accept it. Its much better if you can go as a company to the regulators with a coherent story and a blanked risk assessment and a reason for why that drug should be safe for those patients in that treatment period in that age group, whatever. Its better if they take the dissections and do the work first. Right we will look at the last slide where at the beginning david mentioned he spends a lot of his time heloing clients try in to sort out those situations where etheyhave some sort of positive or negative results and they are trying to tease the real positives forth false positives and the real negatives form the false negatives. One of the big problems is that once you do a test and think abc to the bar charts that we saw is that what criteria do you use to call something positive? And across the literature and across the years there are lots of different ways that people approche it and every body got there favourites. You might use statustical significance, thats probably a good thing for some of the mammalian cells tests but its very sensitive for small increase for example in the Ames test. You might want to use a full increase over your control. You might want to look at your historical control rate and say well something is only positive if it goes out side the historical conrole rate that we have built up over a period of time. Thats good for a low frequency event but probably not so good for things like the Ames test or the mouse lymphoma assay. You might set a preset levels for the mouse lymphoma assay we had a working group that collected data from about 200 labs and they established a control distributetion and said well alright well we need see a umant freequency of X. So if the normal is Y then that diffenree btween Y and X is a mathematical increase that we would never normally expect to see within the controls population so we will put this preset bar where they say anything grater or increased beyondd this point has to be biological significant. They call this the global evalutaion factor. This si the only test system where we know of that this happens. It might however come in to others. There is then dose response where people are more convinced that if people see a dose reponce across several concnetration rather than at a single point. At a single point they don't know what it means and it may be due to toxicity at the highest dose or in the middle of the dose range. What is it telling us and reproducibility. We have mentioned that now some o the regulators are only doing a single experiment. but most people feel much more comfortable knowing they have seen either a positive or a negative that is reportducible. It takes out one of the elements of chance that any experiment can give you a diffenret result on different day, even using the same compounds the same technicians the same glass and plastic wear because these are biological systems

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and they have inbuilt heterogeneity. So these are problems for interpreting results for any body and they are pretty much every body has there choices and there all doing differ ones.

We are just going to spend five minutes introducing the next lecture so that we can scoot through it. So the agin just shows that with in the cancer process there are several step where ells of genetic material is important and so chromosomal damage and thats what this next talk is about (DNA damage, Chromosome damage) now this is distinct form mutations. The chromosome damages do play a significant t role in the



tumor changes, through initiation progression and inheriting diseases. Mutations and other events including chromosome damage generally lead to something that we put in the umbrella term genomic instability. This includes a raft of things like chromosome loss or aneuploidy, abnormal chromosome segregation, deletion of part of the chromosome or recombination between chromosomes, these are all typical of unstable genes. This is why when you look at cancer cells you see lots of genetic changes, those are changes that have occurred as the cancer has grown. They are not what cause the initiation event they are what has happened as a result of the uncontrolled rate of proliferation. Once a cell becomes mutated and once it has experienced some genetic change then the whole genome seems to become unstable and you get a build up of these other kinds of changes. Genomic instability is associated with an increased cancer risk. The best examples that we have are Blooms syndrome, colonic carcinomas and retinoblastoma. These are familial so there is a genetic basis associated with genetic instability but more startling perhaps is that there are more thousands of genetic disorders that exist and many of these are at a very low frequency and you won't have heard of some of them but you will have heard of the most common one which is down syndrome. Which is an additional chromosome 21 there are three copies rather than two. Some of these others we might have heard of are seen below:

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>4000 described. Some are manifest at birth, e.g.:

Down's syndrome (trisomy 21)

Patau's syndrome (trisomy 13)

Edward's syndrome (trisomy 18)

Klinefelter's syndrome (XXY)

Turner's syndrome (XO)

These are all manifested at birth so these are numerical chromosomal changes that are occurring in the reproductive process.

Others manifest themselves later, e.g.

Duchenne muscular dystrophy (childhood)

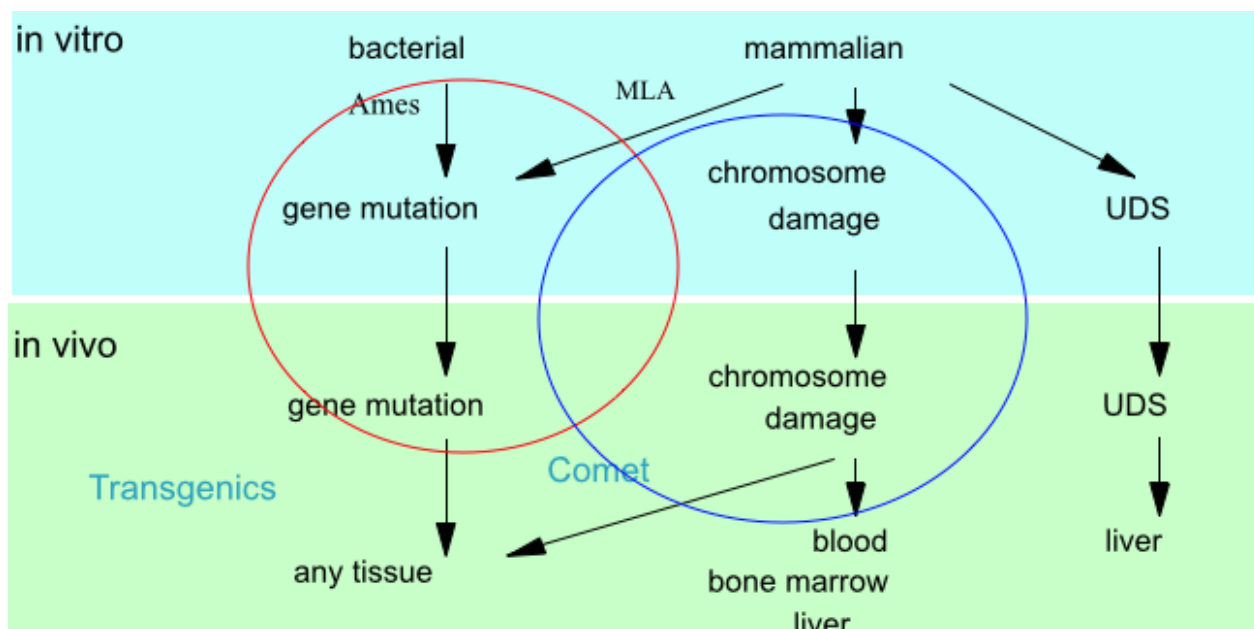
hypogonadism (adolescence)

Huntington's chorea, polycystic kidney disease,,Alzheimer's disease (later in life)

Others only appear later on in life such as huntingtons. So a lot of disease conditions can be associated with chromosomal change. Even more starteling and we are not going to read all of these. You can look at them later. It is estimated, and this goes back to what we were saying a while ago that humans seem to be pretty good at recognizing a genetic abnormality and probably report it at the first month of pregnancy. So we may often not know that even a woman is pregnant. Based on all of the data that we have got we have estimated that 8% of all our embryos formed are chromosomal abnormal. Most of those will not survive and most of those will be aborted but as you saw on the previous slide that some do get through a viable birth. So chromosomal damage is important and as we mentioned in the earlier talks we are not just measuring gene mutations which is what the previous talk was about but we also need to look at structural and numerical chromosome damage because it's important. Not just for the cancer process but for other aspects of health so we have tests that we can do in vitro for chromosomal aberrations or micronucleus and we can include and just get a way with two tests to minimum the effort then an Ames test and an in vitro micronucleus test will pick up both structural and numerical change. As we mentioned before high concentrations and extreme conditions and the need for metabolism. In vivo we can look for chromosome damage but only in limited tissues. We can only really look in bone marrow and blood and we can measure the micronuclei in the liver. So unlike the transgenic mutation assay where we can look at any tissue if we are looking specifically for chromosomal damage then we are limited to only a few tissues and that can be a problem because we need a sensitive assay and that's what we will come on to last this morning. We can look for DNA damage most, chromosomal aberrations result from a double strand DNA break and we can look for those breaks directly using what's called a comet assay and that we can do in any tissue. So we can look directly for chromosomal damage we can't look directly for micronuclei in any tissue but we can look at the precursor DNA double strand breaks in any tissue. So that's what this talk is going to be about. So if we look across in vivo and in vitro we can get gene mutations in vitro and in vivo which we have explained in the previous talk. We can look for chromosomal damage in vitro and in vivo but in vivo it is only in limited tissues and so we have the comet assay that tells us about DNA strand breaks that we can do in any tissue. So it's really just trying to square the circle in terms of looking at all of the relevant endpoints in both in vitro and in vivo. So we

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have already mentioned the important of doing that so we are going to describe three types of assay looking directly for structural chromosome damage looking directly for micronucleus induction and looking for DNA strains brakes.



For structural chromosome damage we can measure this in cultured cells in vitro and in certain tissue, generally bone marrow or blood in vivo. What were doing is actually looking at the chromosomes when they are visible so that when there in the metaphase stage of mitosis or meiosis. So the cells have to be dividing and this is what limits us in terms of the number of tissue we can look at in the whole animal because getting dividing cells is not easy from a tissue like liver or kidney or something of that sort. The bone marrow and there for the cells which are produces in to the blood or the white cells within cells within the blood we can make them divide after treatment we can take blood and stimulate the white cells to divide by using a mitogen like phytohaemagglutinin, that will stimulate the white cells to divide. Then we can look at the chromosomes. Now the problem with scoring chromosomal aberrations is that normal chromosomes at metaphase can take on a number of different appearance. They can look quite different just from one cell to another. So it requitress quite a lot of extensive training, its a bit like pathology but pathologist has to spend a lot of time looking at the way normal cells appear in normal tissues before they can diagnose abnormal cells. Or abnormal tissues and its the same with metaphasee tissue. Before you are aware of all of the different ways that normal chromosome can appear then you cant really identify how abnormal chromosomes. So We will look at some pictures in a minute and it will illustrate that. So basically the cell types that we use fro chromosomal damage. In vitro we can take normal lymphocytes and whole blood. We can stimulate the cells to divide because within the blood they are in a resting stage called called :G0 and we can stimulate those cells to divide by treatment with a mitogen or we can use established cell lines. Most often these are from chinese cell hamsters and that useful because they have only got 20 odd chromosomes so there easy to count. V79, CHO, CHL are the most common . V79 come form lung the CHO are overy and the CHL are also lung. These cells are

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immortal and they grow exponentially but in becoming mortal they have already become chromosomally rearranged. None of these cell lines are truly diploid they have already undergone some chromosomal rearrangements. If you remember what we said before the break. Once a cell loses some of its integrity. Once it starts to become genetically abnormal it tends to become genetically unstable. So further changes occur randomly and spontaneously and that can cause a problem. Also all of these three most commonly used Chinese hamster cell lines are p53 deficient. So if a cell is p53 deficient then it can experience DNA alterations but it won't recognise them and therefore it won't go into apoptosis. So cells that should be dying because of the DNA damage carry on dividing and we see chromosome damage in cells that should be dead. Because they're p53 deficient. and that can lead to misleading positive results as those cells should be dead and there are not. So the latest OECD guidelines state that we are most likely to get reliable results if we use p53 competent human cells such as normal lymphocytes or as we will see when we come to look at human nuclei there is a nice human cell line called TK6 which is p53 competent and that's really quite useful for doing micronucleus test. In vivo we have to use cells that can divide so bone marrow cells and spermatogonial cells or we can take blood from treated animals and we can stimulate those sites ex vivo and we can look at chromosomal damage in them. The normal test protocol is quite similar to that for the mouse lymphoma assay that we discuss before. So the presence and absence of S9. We can't treat for longer than 6 hours because of the toxicity of the S9 but in the chromosome test we almost always include a prolonged treatment between 20-24 hrs so around about one and a half cell cycles in the absence of S9. This is to pick up the sort of compounds that need to be present for a whole cell cycle in order to produce their effects. Classic examples of this are nucleoside analogs things like 5-fluorouracil and things like xeromethyadine need to be present during a division cycle and they need to be present when the DNA is replicating because that's where they're going to be incorporated into DNA and that's when they're going to produce their effects. If you only do a short treatment like for 6 hrs then you might miss that. We sample one and a half cell cycles after the start of treatment which is to allow for any cell cycle delay if the cells pick up damage and try to repair it. In certain circumstances we may actually include a sample time which is 24 hrs later than one and a half cell cycles. So it could be sampling at 20-24 and 48 hrs to pick up any delayed effects. As with the other mammalian cell assays. A number of concentrations preferably due to duplicate cultures but you can use single replicates as long as you score the same total number of cells. Now we mentioned earlier this morning that we don't like zeros in these tests. So to avoid zero counts in the control cultures OECD have recently increased the number of cells scored to 300 cells per concentration. That usually will mean that you're getting real positive numbers of aberrant cells in your control cultures so you can establish the statistical power of the test. Again we are testing to extreme levels where we are going to high concentrations for everything except human pharmaceuticals where they are lower (no one's asked me why they do that by the way). So why is it lower for human pharmaceuticals than for any other type of chemical, ITs because we have much much more safety data and in most cases we will have carcinogenicity data so the regulators are less worried about picking up absolutely everything in the gene to test. This gives you enough data to be comfortable that you are not harming healthy volunteers and patients in the clinical trials. By the time it gets on to market you are going to have a cancer study so if there were any genotoxic effect then it would be picked up there. so they have pushed the limit doesn't allow more compounds to get into clinical trials easily. So we are testing to high concentrations and we

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are testing to relatively high levels of toxicity. We need to see around 50% toxicity in these experience to make sure that we haven't missed anything. By getting into these levels of toxicity you are overwhelming the defence mechanisms of the cell you therefore staining a better chance of any true genetic effects coming through. But you can introduce misleading results and artefacts or misleading positives as result of the toxicity that's occurring in these extreme conditions. So this is a obvious select picture of a human lymphocyte chromosome prep. All of the chromosome are nicely separated and easy to score and all of the chromosome are nicely arranged and there not a lot in the way of kinks or bends or anything like



that.

At 1 we have a complete chromatid interchange and at 2 we have an incomplete chromatid interchange. So these would be the sorts of aberrations that you would look for. We are going to come back to this picture in minute. This is another human lymphocyte cell on the right but look now at the difference in the way the chromosomes appear and you will see what David means about the importance of training. It wouldn't be too difficult to pick up is an aberration in this cell. But in this one the chromatids are much more kinked so are there breaks or is the 1 a break because it looks out of line. What we have here is a double chromatid isochromatid fragment or a deletion. So just think forwards to what will happen to this fragment and what will happen to these interchanges when the cell tries to divide. So we have stopped these cells at this point so that we could view the chromosomes but if we hadn't done that then we would have allowed those cells to go through complete division and go through complete interphase. What would happen to these exchange figures and the fragment in the other cell? Well these might go to one daughter cell but they can't split and go to both because of the rearrangement. The same is true with the fragment because the fragment in this preparation. It hasn't got a centromere so it can't attach to the spindle so it might by chance end up in one of the daughter cells or daughter nuclei, it might not. So there is a potential that this fragment or these exchange figures just get lost and do not appear in either of the daughter nuclei. You'll see the relevance of that in a minute. Just keep that in mind. So that's how we measure chromosomal damage in vitro. In vivo we can do this in bone marrow because bone marrow cells are dividing and we usually do this as an acute study. We can do a single administration and sample at two time points but this uses more animals than doing two administrations and sampling at a single time

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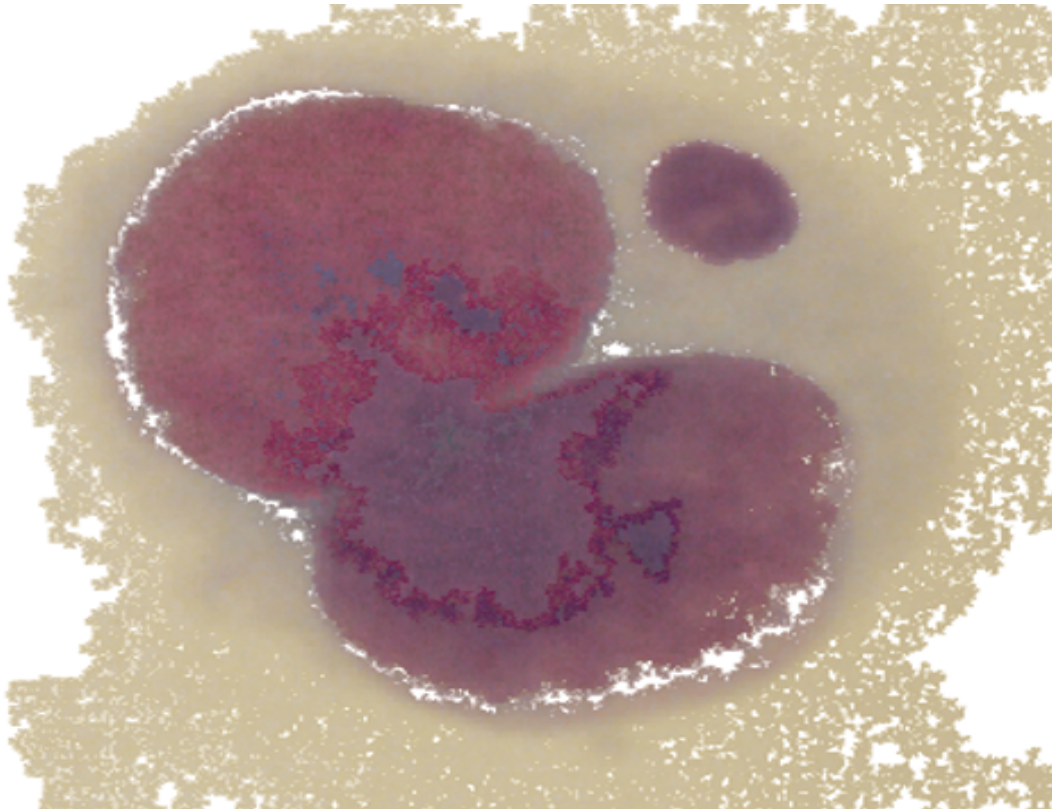
point. Around 12-18 hrs after the last dose around one cell cycle after the last dose. Because we have done two administrations we are allowing for cell cycle delay. Usually five groups of animals per set per group we can do the study in only one sex and therefor reduce the numbers of animals. if there are clearly no sex differences. Again several dose levels and we need to be scoring quite a lot of cells. This has recently been increased, it has been doubled and has been in the latest OECD guidelines again to avoid zeros. So we are not getting zero aberration counts in controls. So this is pretty demanding and you think of the training that goes in to understand all of the different ways that normal cells can appear and then you have to score 200 cells per animal, 5 animals per group and 100 cells per dose group. Wearing down the microscope to see if there are any breaks or fragments or rearrangements. It is very demanding and very time consuming. In these in vivo tests we usually also take blood samples and analyse the amount of test chemical in the blood. This is important because if we get a negative result after scoring all of these cells. We need to know that the bone marrow was exposed. Now there are some compounds that you would administer orally and they would go straight through the GI tract. Come straight out in the feces and nothing gets into the systemic circulation and therefor the bone marrow and the target tissue wouldn't be exposed. So it's not surprising that you get a negative result. The regulators require that we have some proof that the bone marrow and the systemic circulation did contain the test chemical or its metabolites. otherwise a test chemical is not worth the paper it's written on. This can be quite demanding for industry like the chemical industry or the food industry. They don't automatically have bioinformatic methods that allow them to measure test chemicals in biological fluids like blood plasma. So that can be quite demanding and quite expensive just to get that bit of the equation right and if we do measure these concentrations then we are particularly if we think that we might have a no effect level or if we think that we might have a threshold effect and there is a safe level of exposure that we can compare those concentrations in the animals to human concentrations in normal use and in order to do a risk assessment. Questions on chromap's?

How do they test it in the blood, do they use flow cytometry or elisa or something? not for structural aberrations like this no, micronuclei which we will look at next yes, but for chromosomal aberrations no machines accept a metaphase finder which saves the time of scanning the slides for suitable preparations and there are image analysis software that will try to tell you have or not a normal karyotype or an abnormal karyotype but to my knowledge every one of those needs human intervention. A human microscopist or cytogeneticist would have to go back and check whether the machine has identified an aberrant chromosome or not. So you can use image analysis to save some time but for structural chromosome damage, we are not aware that there is an image analysis software out there that can do totally automated scoring with 100% reliability without any human intervention.

Which is why it's so demanding and which is why the micronucleus test which we are going to talk about next has become so popular. These kind of rearranged chromosome and fragments like this may not get incorporated into the daughter nuclei after division at all and if there not then they can end up as a fragment separate from the two daughter nuclei, get wrapped up in a little bit of nuclear membrane and therefor they can look like a micro or small nucleus, we can see these in the interphase cells so after a cell has gone through cell division into the next interphase that's where we would pick up these fragments or

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rearrangements, whole chromosomes as micronuclei. We can do this again in vitro or in vivo using the same cells or the same tissue. Again though we have to have cells that have divided. We can't do this in non-dividing cells but micronuclei are much quicker and easier to score. Do not require the same amount of training, it's much easier to automate and because of that we can score many more cells per sample so we can increase the statistical power of the test. It is important that we know the cells have divided otherwise a negative result or absence of micronuclei isn't telling us anything. It might just be that the cells haven't divided we are looking at interphase nuclei. Cells with an interphase nucleus we don't know whether they have divided or not. So one of the favoured ways to approach this is to use a little trick. We add a chemical called cytochalasin B which blocks cytoplasmic division. So it allows nuclear division but blocks cytoplasmic division therefore we end up with binucleated cells. If we can see binucleated cells we know they have divided. So that gives us a target population that we know has divided in the presence of after the treatment with the chemical and that's important. Again we can use all of the different cell types that are used for chromosomal aberrations but p53 competent human cells are preferred. So it could be for example French blood cultures or it could be what's becoming quite common are these human lymphoblastoid TK cells. These are not tumor cells. They are cells that have been immortalised by treatment by Epstein-Barr virus. So they are not tumor cells as some of the other tumor cell lines that are around like HEPG2 or HeLa. Those are derived from tumours. These are not derived from tumours, they are human lymphoblastoid stem cells that have been made mortal by treatment with Epstein-Barr virus. Similar sort of protocol to the chromosome like the aberration test in terms of numbers of treatments for example. But we sample a little bit later. We need get the cells past that mitosis and into the next interphase. so instead of one to one in a half cell cycles after the treatment, we are going to sample one and a half to two samples after the starter treatment. So this is a binucleated cell seen below:



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With a micronucleus, as we said there is a similar sort of protocol to the chroma's with a number of concentrations preferable duplicate cultures but you can do them in single cultures as long as you score the same total number of cells. If you do use single cultures it's better to use more concentration so you can see the dose response curve. We have routinely scored two thousand cells, so this is ten times more for chromosomal aberrations. 2000 cells per concentration done give us lots of zeros so it's not been increased in the latests OECD revisions. Again we need to get to around 50% toxicity with concentrations for around the same as the chromabs. now we mention that we can get micronuclei both from fragments that haven't got a centromere and from whole chromosomes that have failed to attach to the spindle for some reason. That failure to attach might be because we have had a rearrangement like those chromatid exchanges or it might be because the spindle itself has been damaged. And those micronuclei will contain centromeres and we can distinguish between micro nucleui that contain fragment and micronuclei that contain whole chromosomes by using a pan centromeric probe. Now why is this important. Because if the micronuclei contain whole chromosomes and if that it's a result of spindle damage then the chemical is not attacking the DNA, it's attacking the spindle. It's a non DNA target. The default interpretation of a DNA reactive genotoxin is that there is no threshold. There's no safe level. We actually now know that there are safe examples but the default assumption that there is no threshold. Whereas if the chemical is damaging a non DNA target or if it's damaging the spindle then it will have a threshold and that's important in terms of risk assessment whether or not we can establish a safe level of exposure that humans can experience. So this is just reiterating what we have already said. The default position is that genotoxic agents of genotoxic carcinogens do not have a threshold. This really comes from tradition biology where radiation gives you linear dose response curve that tracks right back to the origin. It's not surprising really because radiation dose does not need a transport system to get into the cell it doesn't need metabolism, it can't be detoxified whereas chemicals need to get into a cell so that's a barrier, they often need to be metabolised to produce a reactive metabolite. So that's another barrier there for there are processes that have thresholds associated with them but the default position is that is assume anything that damages DNA is like radiation and it doesn't have a threshold. So you have got to prove that your chemical has a threshold and therefore it is safe for human exposure if you possibly can. so for chemicals that damage a non DNA target we treat them the same as we would a toxic substance or a non genotoxic carcinogen that is there will be a no effect level. Somewhere below that level then it will be safe to expose human beings because you're not going to see any genotoxic effects low the level at which you see toxic effects. So if we find micronuclei that are centromere positive so it's an effect on the spindle then it's important to establish a no effect level but micronuclei are only one manifestation of aneuploidy. We can get chromosomes that distribute unequally. If there is damage then there is some damage to the spindle we can get chromosomes that distribute unequally to the daughter nuclei. So if it's a human cells there are 47 chromosomes in one nucleus and 45 in the other. There are no micronuclei. None of those chromosomes have been completely lost. But there is an unequal distribution and that is still a health problem. How do we pick that up? that is called non-disjunction when you get an unequal distribution. How do we pick that up? could that occur at concentrations lower than we see micronuclei. Because if it does then the no effect level for non disjunction is more critical and is more sensitive than the no effect level of the micronuclei. So in terms of establishing safety we need the most sensitive measure of aneuploidy and

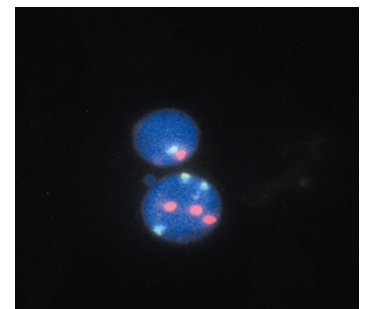
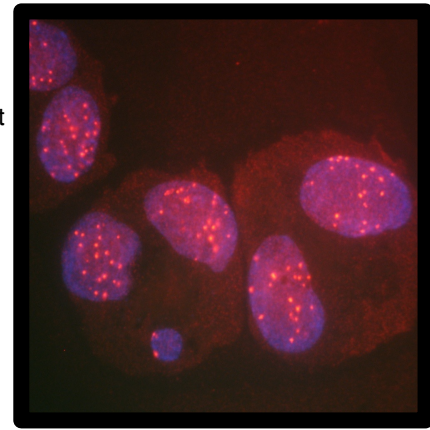
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that would be non-disjunction. We tend to think of the spindle as being a bit like the cable suspension bridge. Those cables on the bridge are not single pieces of metal they are made up of multiple fibres and the spindle is made up of multiple fibres. You could go along to a suspension bridge

with a hack saw and you could cut through one or two of the strands and the bridge would not fall down. But cut through enough of them and it will and it's the same with the spindle. You can induce a certain amount of damage or slight damage to the spindle and there will be no consequence as there are no micronuclei. You could induce a bit more damage and some chromosomes will fail to attach to both sides of the spindle and will segregate abnormally giving you your unbalanced chromosome distribution. Like chromosome 47, 45. So non-disjunction will occur where there is some damage to the spindle but not enough damage to lead to chromosome loss. That will occur at a higher concentration. That's where we get micronuclei and if we go even higher then we create so much damage that everything just falls apart, the cells stop dividing and then the micronucleus level

comes down because there's nothing going into the next interphase. They are all dying at mitosis because they can't complete division. So the important question is we can establish a no effect level for micronuclei and we can establish a no effect level for non-disjunction but how big is that difference and is it important. If we are going to measure non-disjunction then we need to use a chromosome specific probe and not a pan centromeric probe. This is pan centromeric probing so lots and lots of suppressant signals within the cells. That's not good for telling us about unequal distribution of chromosomes. Here's another one including micronuclei with pan centromeric probe. We need to look at whole chromosome probes, so this is a binucleate cell treated with culture seen. We probed here for chromosomes 1 and 8 and this is a human lymphocyte. You see there should be two green and two red signals if the chromosomes have segregated equally and we have got a double non-disjunction. So this is how you measure non-disjunction you use whole chromosome probes for two or three chromosomes. You probe anymore than three and it becomes messy and you can't score it. But that gives you enough that gives you a sensitive measure of non-disjunction. So that's how we can measure non-disjunction. How important is it that we measure it well we now have quite a lot more data and people have done non-disjunction experiments and compared them with micronuclei experiments and the difference between the no effect level for micronuclei and the no effect level for non-disjunction is no more than a factor of two. It's really much closer together than we thought it would be. So what this means is that if you're developing a product you do your micronucleus test. You show by pan centromeric probing that there's mainly centromere positive so they have an aneuploidy effect and there will be a threshold. You can establish a no effect level and you can compare that no effect level with your human exposure. If that margin is very big then you don't need to

- ▶ 3 stage process
 - *In vitro* micronucleus test
 - Pan centromeric probe
 - chromosome specific probing (non-disjunction)
- ▶ Identification of threshold values ("safe level")



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worry about the fact that the no effect levels for non-disjunction may be two fold lower. If this is 1000 fold and it would be 500 fold here it is still massive and nobody cares. But if you're looking at small margins and if this is only ten fold and therefore your non-disjunction non-effect level may only be fivefold then it's going to get an accurate figure. So on occasions even if we have centromere positive micronuclei we have got an aneuploid, we can establish safety margins. We may have to go on and do these non-disjunction tests. To get a more accurate measure of the safety margin. That's micronuclei in vitro.

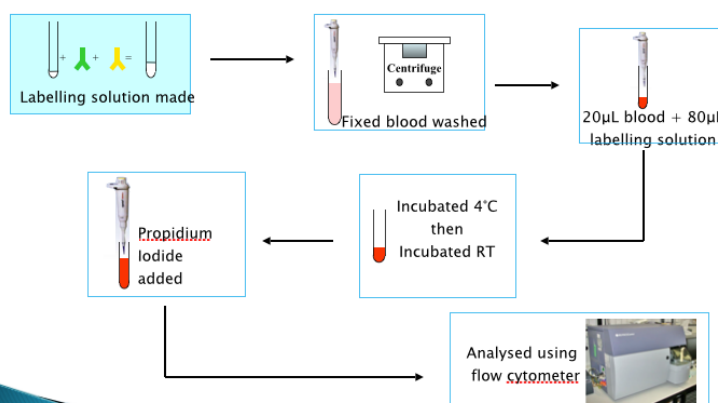
Ok so we can do pretty much the same in vivo. but in vivo we tend to do it in erythrocyte precursor cells any idea why? They don't have a nucleolus which makes it really easy. So we are not having to search for a micronucleus in a cell that contains one or two nucleoli. The erythrocyte once there pushed out into the bone marrow as a young erythrocyte they lose their main nucleus. So if a micronucleus is formed like a fragment or a whole chromosome is formed during the last nuclear division that micronucleus is left in the erythrocyte when the main nucleus is extruded. So it's really easy to score. We can also measure micronuclei in blood. Those young erythrocytes which are called reticulocytes as they still contain RNA until they become fully mature and that RNA is used to synthesize all of the haemoglobin and porphyrins and such that are needed for a mature red blood cell. Those reticulocytes we can score micronuclei in them in the blood. So we can score both in bone marrow when the cells are first formed and in the young reticulocytes when they are pushed out into the blood. We can do the same centromere probing as we did in vitro to look for whole chromosomes to look for aneuploidy. So when the cells divide if there's damage then it will form a micronucleus when the nuclei are extruded. Then that micronucleus gets left behind and we can pick it up in the immature erythrocyte within the bone marrow or in the blood. So we tend to use these days a DNA specific stain like acridine orange. So these big yellow blobs are the nucleated cells. Here are your immature erythrocytes which are red because we do a counterstain with pyrimidine iodide which has got or will stain the RNA. You probably can't see them but there are some really dark almost ghost cells, these are the mature erythrocytes and there won't be many of them in the bone marrow because the maturity mainly happens within the blood so we quite easily distinguish the young erythrocyte and the micronuclei using a stain like acridine orange. So typical study design for a bone marrow test. We can do one administration with two sampling times but this uses twice as many animals so more common is to do two daily administrations and sample 24 hrs after the second dose. We can recombine this micronucleus test with the comet assay. Which we will come onto in a minute and for that we would need to three administrations. This is to incorporate the optimum sampling time for the comet assay so we dose at 0, 24 and 48 hrs and sample three hours later which is what we need to do for the comet assay. So we can combine two endpoints to a single set of animals which is good. We can also do bone marrow and blood micronucleus sampling at the end of a standard 28 day toxicity study. So we don't need to use any additional animals. We can incorporate it in to standard testing. The same sort of recipe where you have 5 animals in 5 groups same sex per group. or one sex if there's no sex differences. Three dose levels if toxic. If non toxic then we might be able to get away with a single dose but if you integrate it into a 28 day study you would have three doses anyway. Recently the recommendations for the number of cells to be scored have doubled again to avoid zeros. So we now score 4000 immature erythrocytes, polychromatic erythrocytes per animal in bone marrow and we use the ratio of immature to mature erythrocytes to give us a measure of toxicity. Again we should measure

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chemical in plasma so we can demonstrate exposure. But we can as we say measure micronuclei in the blood. And for this we can use flow cytometry and for that reason we can score a lot more cells. Now these young reticulocytes, we recognise them in a whole blood sample because they express the CD71 marker, so we can have a fluorescently tagged antibody to CD71 and identify the young reticulocyte population. Now the longer you treat the more likely it is that the spleen will remove a micronucleated erythrocyte from the circulation. To compensate for that we prefer to score larger numbers of cells than we would if we were doing it manual. In fact as long as we are picking up these young reticulocytes they are generally not recognised by the spleen so even if we have got micronuclei they tend not to be removed. But it is easy using flow cytometry to score anything from 20,000 to 2,000,000 cells per sample. So it really gives us good statistical power. And you don't need large blood samples. So you can do cereal sampling you don't need to sacrifice your animals like you do to get at the bone marrow and you can add those cells on to the end of a standard toxicity study and you're not interfering with the tissue that the pathologists want. So it's really neat to be able to include this to give a really good statistical power of information without using additional animals and without interfering with the normal toxicology work. So flow cytometry or FACS is characterisation of single cells as they pass at high speeds through a laser. Light scattered forwards indicates refractive index and this is dependent on cell size, organelles etc. So for example the nucleated cells in the blood will all scatter light forward so they can be gated out so you're not confusing the population that you want to look at. You can gate out the nucleated cells because of the forward light scatter, light scattering side ways indicates density or cell surface granularity. So we can flow the blood samples through the flow cytometer and look at the forward and the side ways light scattering and using our various markers then we can pick up what's happening. This is just a trial when David's lab got involved in this lighttron were the experts located in Rochester New York state and David did some experiments where we treated animals we took the blood. We fixed in this -80 freezers as these blood samples have to be kept very cold which is critical. Because if you don't what happens is that CD71 cell surface marker becomes internalised instead of sticking out from the cell surface where we can identify it with an antibody, it turns inwards and so you can't identify those cells. So this -80 and methanol is absolutely critical for good samples and we just split the samples we need half of them in David's lab and half over to Cytron and they analysed them and that helped establish the method. So you need a labelling solution but you need to pick up these CD71 markers identify the population that you're interested in so you take the fixed blood sample add the labelling solution, incubate, add propidium iodide as a counter stain and run it through the flow cytometer. So we have already gated out the nucleated cells. We have sorted out the cells that are going to be within the gating properties that we are looking at are mature erythrocytes which won't have CD71. We have reticulocytes that will have CD71. Micro nucleated reticulocytes that will have CD71 and will stain with propidium iodide and then platelets which actually

Peripheral Blood MN Methodology

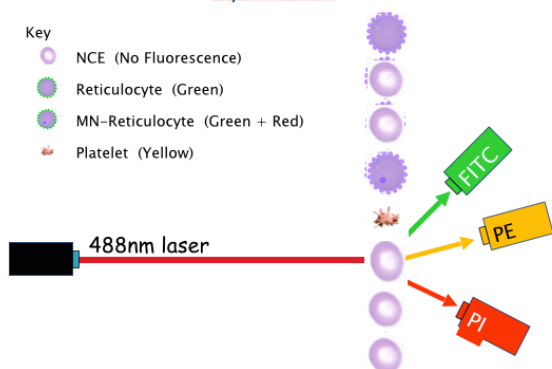
Sample preparation



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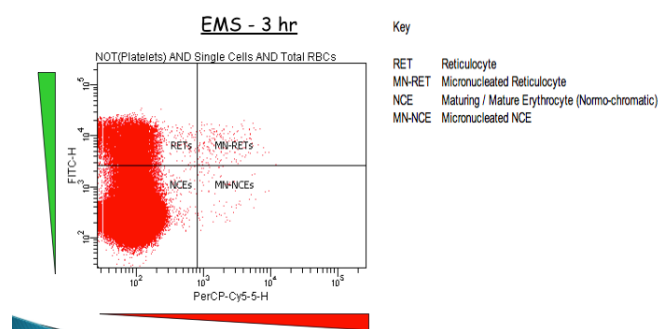
come up yellow because of the stain techniques that were using. so as we pass the blood through the flow cytometer. We get a dingla for a reticulocyte which dosnet contain a micronucleus and then we can get a single for a reticulaocyte so that will give us both FITC and PI. The platelet will only be pickedd up her ebecuase its yellow and so we can get a diatribetion

Peripheral Blood MN Methodology Inside the Cytometer



Peripheral Blood MN Methodology

Data Output



VOX

like the one seen below. Based on the fluorescent reactions that we are getting as the sample goes through the flow cytometer. So this is normal reticulocytes, this is normal mature erythrocytes, micronucleated reticulocytes and micronucleated mature erythrocytes and so we can establish frequencies and quantitative data. So this is for methyl ethyl sulfaphate. Which we would normally have much fewer micronucleated reticulocytes in this box for control sample. We can do micronuclei in control tissues other than bone marrow and blood, there is now a lot of data on measuring micronuclei in the liver. Because you need dividing cells the old method used to be surgical intervention. We used to cut part of the liver away because it regenerates rapidly we could treat the animal whilst the liver is regenerating, but that surgical intervention is demanding, so a group in Japan started looking at young rats. Less than six weeks old where the liver is still dividing because it has not fully grown. and that looked promising but then there were questions regarding the metabolism in a young rat is not the same as the metabolism in a mature rat. Maybe we could be getting misleading results. Recently a trial was done in Japan where they dosed animals, they dosed rats for 14 or 28 days without any surgical intervention or without any additional chemicals and there is enough cell division within the liver during that 14-28 day period that you can effectively measure micronuclei. We are just compiling this into a special year of mutation research which is going to be published soon. So looks as though this is going to be very promising and we will probably end up with an OECD guideline for this in a few years. They also started looking at micronuclei in stomach and colon and again after 14 or 28 days there is enough cell division going on that you can measure micronuclei in those tissues. so it looks as though we may be able to do site of contact micronucleus work as well. So liver as the major organ of metabolism and stomach or colon as the major site of contact for the orally administered substances. looks as though as going to be an option for us going forward. So that's micronuclei. Ten minutes to finish off on strained barking.

So as we mentioned we need dividing cells for micronuclei and compounds as we can't easily do it in any old tissue. So whilst there is some promise that we can do micronuclei in liver and in the GI tract things like

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kindness, spleen lung etc are not easy. So we can take a step back and take a look at the kind of DNA strand breakage that leads to chromosome aberrations and for this we tend to use the

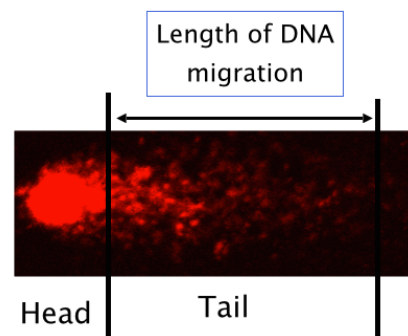
comet assay, so this is a single cell gel electrophoresis assay where we are looking at the migration of broken DNA in an electrical field. All you have to be able to do is create a single cell suspension. So you can look at any tissue from which you can create a viable single cell suspension. This is pretty easy for everything apart from skin. Skin is not easy to convert into a single cell suspension. It takes pretty aggressive digestion to break down all the protective layers starting from the epidermis etc to give you single cells. And it's difficult to get good viability from the skin but pretty much every other tissue you can make a single cell suspension and you can do the comet assay, and cells do not have to be dividing, so that's why it's easy to do in any tissue. We tend to use alkaline conditions because this gives us a wider range of DNA lesions, and we now have last year and OECD guideline for the in vivo comet assay was adapted by OECD, people don't tend to do in vitro comet assay very much, we have got such good mammalian in vitro test anyway that you don't really need to do it, so the emphasis is really on in vivo use of the comet assay. So once we have got our cell suspension we mix it with the cell suspension with agarose, spread them on microscope slides. Gently lyse the cells so the DNA can be released and unwind the DNA so that any fragment can migrate. Carry out electrophoresis under standardised conditions then neutralise, stain and do image analysis which again is usually automated. Unwinding at different pHs expresses different types of damage and the more strand breaks we get the smaller the DNA fragments, the more the migration. So if we were to do the test at neutral pH we would only really pick up double strand breaks and chromosome crosslinks. Not crosslinks because they're difficult to detect because you have to have a high level of migration in your controls and you see a decrease in migration in your treated cultures. So crosslinks should really have a big question mark against it. The standard test is not designed to detect crosslinking agents but you can if you fiddle with the conditions. At alkaline pH we increase the number of genetic damages that we can pick up, so at pH 12 we pick up exact repair sites and above pH 13 we also pick up alkali-labile sites. So in order to get the best possible bang for your buck we tend to do the alkaline, at the greater than pH 13 assay as standard. Why is it called the comet assay because the migration of the DNA on the gel looks like a celestial comet. There are different ways you can measure the extent of the damage, you can look at tail length or you can look at the amount or the percentage of DNA in the tail compared to the total or you do what's called a tail moment which is the amount of migrated DNA multiplied by the tail length. Most

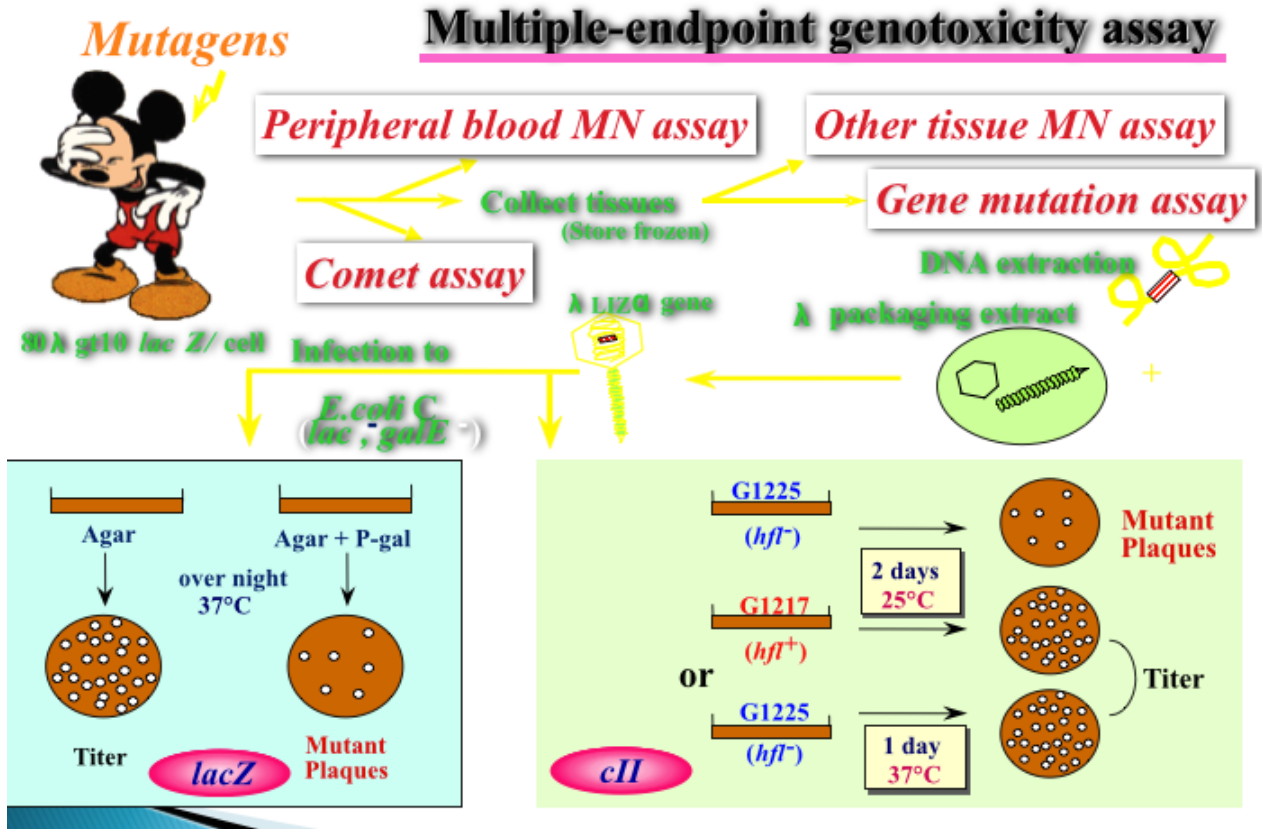
Data Collected by Image Analysis

Cells with DNA migration

Cells without DNA migration

- **Length of DNA migration**
(smallest detectable DNA)
- **% Migrated DNA**
(amount of migrating DNA)
- **Olive Tail Moment**
(migrated DNA x tail length)





people use the percent DNA in the tail so the percent of Dna in the trial, the percent of migrated DNA, that is the most widely used and widely recommended perimeter. Controle cells with very little DNA migration, cells treated with positives ccontrol chemicals have lots of migration. This was just for a bit of fun where ea college of david kurklands in japan did a few years ago said we dont need to do all of these in vivo test. all we need is one super animal and we can do anything. We can do micornuclei in blood, we can do micronuclei in other tissues, if we do this study in a transgenic animal then we can do mutation as well. So lets do everything in one animal. Ets have a super micky and we can do all of these end point in a single animal. Which may happen one day and we a re moving a little bit towards it but were not quite there yet. So again as divd mentionedd at the ned of the first talk all of this data generation is very intersting but there are still a lot of diffenret way in which people will cconsider wether the deathly have got in formunt of them is postive or not. Some prefer statistical analysis. Some prefer a fold increase over control. others look t the historical control range and other will perhaps use a preset level as we do fofr the mouse lymphoma assay. Dose response is important, if you do the experient more than once then reproduceability is important anw what we can see is that this is how it used to be and we have had several battles with OECD to try and get some better thinking on this and in the latests guildnes to be approved we have come up with a multiporngedapprooch that people seem to buy into. And what we have said is ask three or four questions of your data set.

1) is R any point statistically significant.

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- 2) do any of the point exceed the historical control range (and we have had to define very carefully how a historical control range would be established)
- 3) Is there is a dose response
- 4) and if you did more than one experiment are the results reproducible.

So its statistical significance, historical control range, dose response and reproducibility. you could substitute the pre set levels for historical control range if there is a good basis for it, and what we have said is, if you answer yes to all of those question and your data is statistically significant, if it exceed the historical control range or a pre set level, it has a dose response and is reproducible then that is clearly positive. IF you answer no to all of these then it is clearly negative. Anything in between is going to be equivocal and it is going to need further investigation. You will do more experiments where you change the conditions, you increase the number of cells you score you increase the number of sampling times or something like that or you move to different test system and look at different end points. So if you answer yes to some but no to others and it is neither clearly positive nor negative, then do more work is basically what it comes down too. But at least we now have some broad consensus over the way to try to conclude on the data that is in front of us for all these different tests and the common approach that we can use for every test system. It doesn't matter if its the Ames test or a comet assay or a micronucleus test we can apply the same criteria to every test system and that at least gives us some consistency. so hopefully going forwards there will be more agreement over the way to interpret these tests.